

World Guide for Drug Use and Pharmacogenomics

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Medical knowledge is constantly changing. As new information becomes available, changes in treatment, procedures, equipment and the use of drugs and novel biomarkers become necessary.

Drug and gene nomenclature has been respected according to international standards (National Center for Biotechnology Information; Human Cytochrome P450 (CYP) Allele Nomenclature Committee). Original authors are reflected in references on the website and attached CD.

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World Guide for Drug Use and Pharmacogenomics

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The Path to Pharmacogenomics

These are times of change and reconceptualization in medical science, education, politics and economics. Healthcare expenditure is becoming a major political and economic problem in the developed world, especially in the USA and the EU. In some European countries, the cost of healthcare is causing a global bankruptcy in the public sector. Pharmaceutical expenditure represents about 10-20% of total healthcare costs. Spending on pharmaceuticals throughout the OECD countries has increased by over 35% since 2000. In 2003, spending on drugs represented around 18% of total health expenses in OECD countries (30% in the Slovak Republic, Korea and Hungary, and 10% in Denmark and Norway). However, a combination of slower growth in healthcare spending and in the expansion of economies has led to a stabilization of health expenditure as a proportion of the GDP in many OECD countries. During the first half of the past decade until 2006, health spending in OECD countries grew in real terms by over 3% on average, the lowest rate since 1997. Health expenditure grew rapidly in many countries between 2000 and 2003, with an annual average growth rate of 6.2%. Since 2003, the rise in health expenditure has slowed to an average of 3.6% per year. In 2006, pharmaceutical spending in OECD countries increased by 2%, compared with growth rates of 6-7% annually between 2000 and 2003, and 3-3.5% per year during 2004 and 2005. The health expenditure share of the GDP in OECD countries remained unchanged, this being between 6-9%. Public spending on prescription drugs in the USA increased by 30% in 2006, because of the introduction of the new Medicare drug program for the elderly and the disabled. Private spending was reduced by 4%. In 2010, the growth in spending on prescription drugs in the USA (\$307.4 billion) slowed to 2.3%, the second lowest level in half a century (1.8% in 2008), and was 5.1% lower than the growth recorded in 2009. When adjusted by the GDP price deflator, real total spending increased from \$261 billion in 2006 to \$277 billion in 2010. Real spending per capita increased from \$876 in 2006 to \$898 in 2010, the highest in the world, followed by Canada, Belgium and France, and at a great distance by other countries such as Mexico, Poland, Denmark and New Zealand, with spending ranging from \$180 to \$300 per capita. OECD pharmaceutical spending was over \$650 PPP in 2010. These international differences in drug expenditure, in part, also reflect similar differences in applied health technology, the furtherance of science and scientific production, together broadening the inequalities among countries and enhancing the economic disequilibrium aggravated by the global crisis in which the engines of the world economy (USA, EU, Japan) are now immersed. This circumstance will undoubtedly affect the implementation and/or maintenance of national healthcare programs, the cost of drugs, new investments in drug development and budgetary decisions in medical science.

For drugs entering human clinical trials for the first time between 1989 and 2002, the estimated cost per new drug ranged from \$500 million to over \$2,000 million, depending on the pharmaceutical category and the company involved in the development of the drug. This kind of investment is affordable by only a few companies. To increase capitalization and accessibility to bigger markets, middle- and small-size companies are forced to undergo merging procedures with the consequent loss of jobs and industrial shrinkage. The pharmaceutical industry has to rebuild its strategic plans to improve its portfolio, and has to face restrictions in healthcare costs, and in the price of medicines influenced by the expiration of patents and by the impact of generics on nominal accounts, together with the challenges posed by pharmaceutical biotechnology and the increasing influence of biopharma on the sector, and the conceptual revolution driven by Genomic Medicine.

The Challenges of Genomic Medicine and Pharmacogenomics

The heroic task of the pioneering leaders of the Human Genome, who were able to motivate politicians and investors to create international consortia in order to accelerate the mapping of genes integrated in the human genome, has crystallized into the conceptual revolution which substantiates the foundations of a modern, innovative genomic medicine. At present, genomic medicine is not fulfilling the naive expectations created 20 years ago; however, the future of genomic medicine will be brilliant and indispensable for the progress of medical praxis in the short- and medium-terms. Health priorities have changed in Western countries. A clear shift from acute diseases (e.g. infections) to chronic diseases (e.g. degenerative disorders) has been observed over the past 50 years in developed countries. In developing countries, infectious diseases still remain as a dominant concern; however, the aging of the population is also modifying the medical epidemiology of developing economies. In countries with low and middle income, dementia makes the largest contribution to disability with a median population-attributable prevalence fraction of 25.1%, followed by stroke (11.4%), limb impairment (10.5%), arthritis (9.9%), depression (8.3%), eyesight problems (6.8%) and gastrointestinal impairments (6.5%).

The major causes of death in developed countries are cardiovascular disorders (25-30%), cancer (20-25%), and CNS disorders (10-15%). According to the NCBI PubMed Database, 2.5 million references on cancer have been published since 1818 (415,371 references on genetics of cancer), 1.6 million references on cardiovascular disorders have been published since 1927 (103,470 on genetics of cardiovascular disorders), and 1.01 million references have been published on CNS disorders since 1893 (110,426 references on genetics of CNS disorders). The growth of the elderly population is a common phenomenon in both developed and developing countries, bringing about future challenges in terms of health policy and disability rates. In the USA, death rates for the leading causes of death are heart disease (200.2 per 100,000), cancer (180.7 per 100,000), and stroke (43.6 per 100,000). Dementia is becoming a major problem of health, and concomitant pathologies are highly common among patients with dementia. Approximately 20% of patients with Alzheimer's disease are hypertensive, 25% are diabetic, 50% are hypercholesterolemic, and 23% are hypertriglyceridemic. Over 25% of the patients exhibit high GGT activity, 5-10% show anemic conditions, 30-50% show an abnormal cerebrovascular function characterized by poor brain perfusion, and over 60% have an abnormal electroencephalographic pattern. Significant differences are currently seen between females and males, indicating the effect of gender on the phenotypic expression of the disease. Elderly patients may take 6-12 different drugs per day, with the consequent risk for adverse drug reactions and side-effects associated with drug interactions.

CNS disorders represent a typical paradigm of healthcare inefficiency in most countries. The total annual cost of brain disorders in Europe is about €386 billion, with €135 billion of direct medical expenditures (€78 billion, inpatients; €45 billion, outpatients; €13 billion, pharmacological treatment), €179 billion of indirect costs (lost workdays, productivity loss, permanent disability), and €72 billion of direct non-medical costs. Mental disorders represent €240 billion (62% of the total cost, excluding dementia), followed by neurological diseases (€84 billion, 22%). Despite the personal impact of brain disorders on those who suffer a deterioration of their mental activities, these diseases pose several challenges to our society and the scientific community: (i) they represent an epidemiological problem and a socio-economic, psychological and family burden; (ii) most of them have an obscure/complex pathogenesis; (iii) their diagnosis is not easy and lacks specific biomarkers; and (iv) their treatment is difficult and not cost-effective.

The major challenges of genomic medicine are concentrated around 4 main areas: (i) pathogenesis, (ii) diagnosis, (iii) treatment and (iv) prevention. The pathogenic mechanisms underlying human diseases are largely unknown; our understanding of major complex disorders is deficient because the pathogenesis of these diseases is poorly defined in more than 80% of the cases. At present, we lack predictive markers of disease (many years before the onset of the disease); and medical diagnosis is based on clinical symptoms (phenotypes) devoid of accurate biomarkers (except for infectious diseases and a few other medical conditions). In many cases, diagnosis is late and inaccurate. In global terms, pharmacological treatment is effective in only 20-30% of the cases. It is expected that genomic medicine will help us to develop predictive markers for an early diagnosis, assuming that in over 70-80% of complex disorders the clinical phenotype is the result of a defective genomic background in conjunction with environmental factors and epigenetic phenomena. Without predictive biomarkers, prevention is a difficult task, if not impossible. There is an appealing golden rule in complex genomics of human disease: the greater the number of defective genes associated with a particular disease, the earlier the onset of the disease and the poorer the therapeutic outcome; and the lower the number of genes affected, the later the onset and the better the therapeutic response to conventional therapeutics. Since treatment and the search for health conditions is the final goal of any medical process, it is reasonable to predict that one of the first priorities of genomic medicine might be the implementation of programs for a personalized treatment. Major impact factors associated with drug efficacy and safety include: (i) the mechanisms of action of drugs, (ii) drug-specific adverse reactions, (iii) drug-drug interactions, (iv) nutritional factors, (v) vascular factors, (vi) social factors and (vii) genomic factors (nutrigenetics, nutrigenomics, pharmacogenetics, pharmacogenomics). Among genomic factors, nutrigenetics/nutrigenomics and pharmacogenetics/pharmacogenomics may account for more than 80% of efficacy-safety outcomes in current therapeutics.

To achieve substantial goals, genomic medicine has to rely on structural genomics, transcriptomics, functional genomics, proteomics, epigenetics, and metabolomics, together with bioinformatics and high-tech diagnostic tools. Structural genomics would define the cartographic disposition and polymorphic variation of genes associated with human disease. Over 12,000 of the 35,000 genes which integrate the human genome might be assigned to specific traits; either Mendelian or susceptibility traits. Functional genomics would show the influence of genes on disease pathogenesis and phenotype expression. Transcriptomics and proteomics would help to elucidate the role of abnormal gene expression in the pathogenesis of a particular disease. The study of genotype-phenotype correlations is essential for the evaluation of the actual impact of specific polymorphic variants of a particular gene on the clinical manifestation of a disease and/or biological markers reflecting the disease condition or different biological states of the individual. Epigenetics is the study of how the environment can affect the genome of the individual during his or her development as well as the development of his/her descendants, all without changing the DNA sequence, but inducing modifications in gene expression through DNA methylation-demethylation or through modification of histones by processes of methylation, deacetylation, and phosphorylation. More than 150 post-translational modifications of histones have been reported, including methylations, acetylations, ubiquitinations, SUMOylations and phosphorylations. Epigenetic changes are probably involved in the pathogenesis of many complex disorders. The metabolome is the repertoire of biochemicals present in cells, tissues, and body fluids, whose dysfunctional interactions may lead to disease pathogenesis. Mitochondrial DNA abnormalities and aberrant interactions between mtDNA and nuclear DNA products are also subtle fields to be investigated in genomic medicine with repercussions in human disease.

Our understanding of the pathophysiology of complex disorders (cardiovascular disorders, cancer, or CNS disorders, representing 60-80% of the major causes of death) has advanced dramatically over the last 30 years, especially in terms of their molecular pathogenesis and genetics. The drug treatment of complex disorders has also made remarkable strides, with the introduction of many new drugs. Improvement in terms of clinical outcome, however, has fallen short of expectations, with up to one third of the patients continuing to experience clinical relapse or unacceptable medication-related side-effects in spite of efforts to identify optimal treatment regimens with one or more drugs. Potential reasons to explain this historical setback might be that: (i) the molecular pathology of most complex disorders is still poorly understood; (ii) drug targets are inappropriate, not fitting into the real etiology of the disease; (iii) most treatments are symptomatic, but not anti-pathogenic; (iv) the genetic component of most complex disorders is poorly defined; and (v) the understanding of genome-drug interactions is very limited.

The optimization of therapeutics requires the establishment of new postulates regarding (i) novel strategies for drug development (a reduction in the time to reach the market, identification of optimal targets, pharmacogenetic-oriented methodologies), (ii) the cost of medicines, (iii) the assessment of protocols for multifactorial treatment in chronic disorders, (iii) the implementation of novel therapeutics addressing causative factors, and (iv) the setting-up of pharmacogenomic strategies for drug development and drugs in current use.

Pharmacogenetics/Pharmacogenomics relates to the application of genomic technologies, such as genotyping, gene sequencing, gene expression, genetic epidemiology, transcriptomics, proteomics, metabolomics and bioinformatics, to drugs in clinical development and on the market, applying the large-scale systematic approaches of genomics to speed up the discovery of drug response markers, whether they act at the level of drug target, drug metabolism, or disease pathways.

The potential implications of pharmacogenomics in clinical trials and molecular therapeutics are that a particular disease could be treated according to genomic and biological markers, selecting medications and diseases which are optimized for individual patients or clusters of patients with a similar genomic profile. For many medications, interindividual differences are mainly due to SNPs in genes encoding drug-metabolizing enzymes, drug transporters, and/or drug targets (e.g. genome-related defective enzymes, receptors and proteins, which alter metabolic pathways leading to disease phenotype expression).

The pharmacogenomic outcome depends upon many different determinant factors including (i) genomic profile, (ii) disease phenotype, (iii) concomitant pathology, (iv) genotype-phenotype correlations, (v) nutritional conditions, (vi) age and gender, (vii) pharmacological profile of the drugs, (viii) drug-drug interactions, (ix) gene expression profile, (x) transcriptomic cascade, (xi) proteomic profile and (xii) metabolomic networking. The dissection and further integration of all these factors is of paramount importance for the assessment of the pharmacogenomic outcome in terms of safety and efficacy. Pharmacogenomic approaches based on genome-wide sets of SNPs associated with drug response are now feasible and may offer the potential to personalize therapeutics.

In recent times, significant advances have expedited the introduction of pharmacogenomic approaches in drug development and also in clinical practice to optimize therapeutics; however, pharmacogenomics is still in a very primitive stage. Personalized therapeutics requires a better understanding of the effects of drugs on gene expression in pathogenic cascades, and the influence that different genes and their products exert on the fate of drugs before, during and after impacting with pathogenic targets. At least 5 different categories of genes are potentially involved in the pharmacogenomic process: (i) genes associated with disease pathogenesis, (ii) genes associated with the mechanism of action of a particular drug (e.g. synthesizing enzymes, catabolizing enzymes, receptor genes), (iii) genes associated with phase I (CYPs) and phase II reactions (UGTs, GSTs, SULTs, NATs), (iv) genes associated with transporters (ABCs, OATs), and (v) pleiotropic genes involved in multifaceted cascades and/or genes associated with concomitant pathologies. On average, 8-10 genes (range: 1-131) may be involved in the therapeutic outcome associated with the efficacy and safety of a specific drug.

The vast majority (78%) of the 200 most prescribed drugs in the USA are metabolized via enzymes of the cytochrome P450 family (CYPs), with major contributions from CYP3A4/5 (37% of drugs) followed by CYP2C9 (17%), CYP2D6 (15%), CYP2C19 (10%), CYP1A2 (9%), CYP2C8 (6%), and CYP2B6 (4%). The genes encoding CYP2D6, CYP2C19, CYP2C9, and CYP3A4/5 isoenzymes are highly polymorphic, with great allelic variation in different ethnic groups. These genes are very promiscuous, influencing the metabolism of different drugs. Very few drugs use an exclusive metabolic pathway for their degradation and subsequent elimination. About 10,600 genes (of the 5 categories previously referred) may participate in the metabolism of 1,392 drugs approved by the FDA, EMEA, and Koseisho. CYP3A4/5 participates in the metabolism of 525 drugs (37.71%), CYP2D6 in 315 (22.63%), CYP2C9 in 287 (20.62%), CYP2C19 in 244 (17.53%), and ABCB1 in 272 (19.54%). Of 7,742 chemicals studied, 982 drugs have been related to CYP2D6 metabolism, of which 205 drugs have been characterized as major substrates, 166 as minor substrates, 75 as strong inhibitors, 183 as moderate inhibitors, 117 as weak inhibitors, and 18 drugs as inducers; 691 drugs have been related to CYP2C9 metabolism (177 major substrates, 134 minor substrates, 102 strong inhibitors, 181 moderate inhibitors, 92 weak inhibitors, and 41 inducers); 576 drugs have been related to CYP2C19 metabolism (151 major substrates, 130 minor substrates, 64 strong inhibitors, 127 moderate inhibitors, 72 weak inhibitors, and 23 inducers); and 1,937 drugs have been related to CYP3A4/5 metabolism (897 major substrates, 136 minor substrates, 141 strong inhibitors, 437 moderate inhibitors, 118 weak inhibitors, and 241 inducers). ABCB1 is a very important transporter, with 1,214 related drugs (897 major substrates, 136 minor substrates, 141 strong inhibitors, 437 moderate inhibitors, 118 weak inhibitors, and 241 inducers).

Although studies illustrating the influence of CYP enzymes on drug metabolism have been reported since the 1950s, our molecular knowledge of CYP pharmacogenetics is still very limited, with many black holes. Some examples are (i) tissue-specific metabolism (CYP expression in different tissues), (ii) age- and sex-related changes in drug metabolism, (iii) blood-brain barrier and trans-placental transport of drugs, (iv) fetal metabolism, (v) the dissociation of *in vivo* and *in vitro* results, (vi) the specificity of ligands for phenotyping studies, (vii) the influence of food in drug metabolism, (viii) the role of nuclear receptors, CYP450 oxidoreductase and cytochrome b5, in the efficiency of CYP isoenzymes, (ix) ethnic differences, and (x) drug resistance.

Numerous xenobiotics are not necessarily toxic as such, but are enzymatically transformed in the liver or in other biological organs, where CYP enzymes are expressed, to reactive and toxic intermediates. Many proteratogens, procarcinogens and promutagens are metabolically transformed by CYP enzymes into dangerous compounds with potential deleterious consequences for human health.

Age, sex, and genetic forms of CYP expression constitute relevant determinants of interindividual variability in CYP-dependent metabolism during ontogeny and throughout the life-span.

Important aspects of drug pharmacokinetics beyond genomic variants still awaiting elucidation are the effects of cytochrome b5 on catalytic activities of CYPs co-expressed with NADPH-CYP450 oxidoreductase. Mutations in CYP450 oxidoreductase can lead to reduced activities of steroidogenic P450s CYP17A1, CYP19A1, and CYP21A1. Mutations in the FMN binding domain of CYP450 oxidoreductase may reduce CYP3A4 activity, affecting drug and steroid metabolism.

Food-drug interactions can be associated with alterations in the pharmacokinetic and pharmacodynamic profile of drugs with clinical implications. In recent years, there has been an increasing application of herbal medicines and dietary supplements to treat chronic diseases and promote health. Organ toxicities associated with consumption of herbal medicines have been reported. Natural products undergoing metabolic activation may form reactive metabolites which can induce toxicity. Some herbal components may also be converted to toxic or mutagenic and carcinogenic metabolites by CYPs and Phase II conjugating enzymes. Numerous interactions between herbal medicines and commercial drugs have been documented. Over 50% of commercial ethanolic herbal extracts can cause inhibition of CYP3A4 metabolite formation. This phenomenon is extensive to all continents where herbal constituents are currently used in folk medicine and natural preparations.

Multidrug resistance is a major obstacle to successful cancer treatment. An important mechanism by which oncogenic cells may become resistant to chemotherapy is the expression of ABC transporters that use the energy of ATP hydrolysis to transport a wide variety of substrates across the cell membrane.

Clinicians currently have no way of predicting who will respond appropriately to a given drug, and the paradigm of trial-and-error is especially distressing for patients with chronic disorders and/or severe acute illness. Pharmacogenetic association studies may provide insight into which genetic polymorphisms might be clinically relevant for personalizing pharmacotherapeutic regimens. In the Western population, only 25% of its members are extensive metabolizers (EM) for the trigenic cluster integrated by CYPs 2D6+2C19+2C9, the most relevant genes (and enzyme products) involved in drug metabolism, together with CYP3A4/5. The other 75% of the population is potentially at risk for developing adverse drug events due to defective variants encoding deficient enzymes which give rise to intermediate (IM), poor (PM) or ultra-rapid metabolizers (UM). This population cluster of defective metabolizers requires dose adjustment to avoid side-effects and, in the case of PMs, the administration of another drug with alternative metabolic pathways should be the norm in order to avoid toxicity.

Ethnic differences must be taken into consideration when developing new drugs or when prescribing drugs which were tested in different ethnic groups. Although global clinical trials can enable the development of new agents efficiently, whether the results of clinical trials performed in one population can be fully extrapolated to another population remains highly questionable due to clear ethnic differences associated with genotype-related drug metabolism. Therefore, pharmacogenomic differences associated with individual responses to drugs should be carefully considered when conducting clinical trials or when prescribing drugs for chronic disorders.

To achieve a mature discipline of pharmacogenomics it would be convenient to accelerate the following processes: (i) educating physicians and the public on the use of genetic/genomic screening in the daily clinical practice; (ii) the standardization of genetic testing for major categories of drugs; (iii) the validation of pharmacogenomic procedures according to drug category and pathology; (iv) the regulation of ethical, social, and economic issues; and (v) the incorporation of pharmacogenomic procedures to both drugs in development and drugs on the market, in order to optimize therapeutics.

The World Guide of Pharmacogenomics

The World Guide for Drug Use and Pharmacogenomics (WGPGx) is a multidisciplinary, systematic exercise to put in order a myriad of data organized in selected databases and thousands of reports scattered throughout the international literature on genetics, genomics, pharmacology, drug metabolism, therapeutics, and pharmacogenomics, to provide the reader (physicians, geneticists, pharmacists, researchers, health professionals, regulators, etc.) with a body of practical information which is not available in other publications as a whole.

The Guide is divided into 5 main parts: (i) Drugs, (ii) Genes, (iii) References, (iv) Appendix and (v) Index.

The section of Drugs includes 1,395 drugs classified in alphabetical order from A to Z. Each entry contains the following headings: Drug Name; Brand Names [in 27 European countries (Austria, Belgium, Bulgaria, Cyprus, Czech Republic, Denmark, Estonia, Finland, France, Germany, Greece, Hungary, Ireland, Italy, Latvia, Lithuania, Luxembourg, Malta, Netherlands, Poland, Portugal, Romania, Slovakia, Slovenia, Spain, Sweden, UK), North America (Canada and USA), Latin America (Argentina, Brazil, Mexico), and Asia (Japan)]; Drug Combinations; Chemistry; Pharmacological Category; Mechanism of Action; Therapeutic Use; Unlabeled Use; Pregnancy and Lactation Implications; Contraindications; Warnings and Precautions; Adverse Reactions (cardiovascular, central nervous system, dermatologic, endocrine and metabolic, gastrointestinal, genitourinary, hematologic, hepatic, local, neuromuscular and skeletal, renal, respiratory, miscellaneous); Pharmacogenetics (genotypes of risk, genes involved in drug metabolism, condition of substrate, inhibitor or inducer, when appropriate); Drug Interactions; Nutrition/Nutraaceutical Interactions; Dosage; Pharmacokinetics and Pharmacodynamics (absorption, distribution, protein binding, metabolism, bioavailability, half-life, time to peak, excretion); Special Considerations (diet, age, sex, monitoring parameters).

The section of Genes includes 447 genes of relevance in pharmacogenomics. Each entry contains the following headings: Gene Name; Alternative Names; Alternative Symbols; Locus; Codes (OMIM, PharmGKB); Gene Structure; RNA; Protein; Function; Related Diseases (list of diseases investigated; in bold face those diseases potentially associated); Related Drugs (list of drugs investigated; when appropriate, major substrates, minor substrates, strong inhibitors, moderate inhibitors, weak inhibitors, and inducers); Animal Models; Allelic Variants; Selected SNPs; Evolution; Genomics and Pharmacogenomics (description of relevant findings in genomics and pharmacogenomics according to the international literature); Drug-Gene Interactions.

The section of References includes two categories: (i) Websites and International Databases, and (ii) 17,947 References alphabetically classified by author and by gene. The List of References is included in the attached CD-ROM. Due to the length of said list of references, it has been omitted in the book. The Guide's authors have respected the data and opinions of the referenced original authors where contradictory results on drugs or genes are reported. Data included in the WGPGx have been strictly extracted from the referenced authors, avoiding personal interpretations. Approximately 100,000 references have been reviewed, of which 17,947 were finally selected for this edition.

The Appendix includes 4 sub-sections: (i) Classification of Drugs, (ii) Genes-Diseases (a list of genes associated with specific diseases), (iii) Diseases-Genes (a list of diseases associated with specific genes), and (iv) Pharmacogenomic synopsis (a list of internationally-used drugs with those genes potentially involved in the metabolism of each drug).

The Index contains approximately 52,000 entries divided into 5 sections: (i) Drugs (approx. 7,750 entries), (ii) Brand names (approx. 31,750 entries), (iii) Pharmacological categories (1,891 entries), (iv) Genes (approx. 4,450 entries), and (v) Diseases (approx. 9,200 entries).

As Editor-in-Chief of the 1st Edition of the **World Guide for Drug Use and Pharmacogenomics**, in 3 formats (Book, CD-ROM, and EuroPharmaGenics Database), it is a pleasure for me to present this monumental work to the scientific and medical communities from whom I would greatly appreciate to receive any feedback and constructive criticism in order to improve the quality and accuracy of the information released in this volume. I would like to thank all referenced authors and the entities responsible for the databases from which we necessarily obtained the documentation bulk, later to be assembled in this Guide. The final goal of this incipient, extensive, and systematic work is to efficiently serve our patients, by improving diagnostic procedures with the aid of genomic tools, personalizing treatments and minimizing undesirable drug effects by using pharmacogenomic strategies, and, in the end, optimizing therapeutics for a better healthcare model in our society.

Ramón Cacabelos, M.D., Ph.D., D.M.Sc.
Editor-in-Chief

Contents

Preface	v
---------	---

Contents	vii
----------	-----

Drugs

A			
Abacavir	1	Ampicillin	39
Abacavir and Lamivudine	1	Amprenavir	40
Abacavir, Lamivudine, and Zidovudine	2	Amyl Nitrite	41
Abarelix	2	Anagrelide	41
Abatacept	3	Anakinra	41
Abciximab	3	Anapso (Polypodium leucotomos L)	42
Acamprosate	4	Anastrozole	42
Acarbose	4	Anidulafungin	43
Acebutolol	5	Anti-Inhibitor Coagulant Complex	43
Aceclofenac	5	Antihemophilic Factor/von Willebrand Factor Complex (Human)	43
Acenocoumarol	6	Antihemophilic Factor	44
Acetaminophen (Paracetamol)	6	Antithrombin III	44
Acetazolamide	8	Antithymocyte Globulin	45
Acetohydroxamic Acid	8	Apomorphine	45
Acetylcholine	9	Apraclonidine	46
Acetylcysteine	9	Aprepitant	46
Acitretin	10	Aprotinin	47
Acyclovir	10	Arformoterol	47
Adalimumab	11	Argatroban	48
Adefovir	11	Arginine	48
Adenosine	12	Aripiprazole	49
Agalsidase Alpha	13	Armodafinil	49
Agalsidase Beta	13	Arsenic Trioxide	50
Agomelatine	13	Artemether and Lumefantrine	50
Albendazole	14	Ascorbic Acid (Vitamin C)	51
Albumin	14	Asenapine	52
Albuterol (Salbutamol)	15	Asparaginase	52
Alclometasone	16	Aspirin (Acetylsalicylic Acid)	53
Alcohol (Ethyl)	16	Astemizole	54
Aldesleukin	16	Atazanavir	55
Alefacept	17	Atenolol	55
Alemtizumab	17	Atomoxetine	56
Alendronate	18	Atorvastatin	57
Alfentanil	19	Atovaquone	58
Alfuzosin	19	Atracurium	58
Alglucerase	20	Atropine	59
Alglucosidase Alpha	20	Attapulgite	59
Aliskiren	21	Auranofin	60
Allopurinol	21	Azacitidine	60
Almotriptan	22	Azathioprine	61
Alosetron	22	Azelaic Acid	61
Alpha1-Proteinase Inhibitor	23	Azelastine	62
Alprazolam	23	Azithromycin	62
Alprostadil	24	Aztreonam	63
Alteplase	24		
Altretamine	25	B	
Aluminum Hydroxide	25	Bacitracin	65
Alvimopan	26	Baclofen	65
Amantadine	26	Balanced Salt Solution	66
Ambenonium	27	Balsalazide	66
Ambrisentan	27	Barium	66
Amifampridine	28	Basiliximab	67
Amifostine	28	Bazedoxifene	67
Amikacin	29	BCG Vaccine	68
Amiloride	29	Becaplermin	68
Aminocaproic Acid	30	Bedomethasone	69
Aminoglutethimide	30	Belladonna (<i>Atropa belladonna</i> L.)	69
Aminolevulinic Acid	31	Benazepril	70
Aminophylline	31	Bendamustine	71
Aminosalicyclic Acid	32	Benperidol	71
Amiodarone	32	Benserazide and Levodopa	72
Amitriptyline	33	Benzocaine	73
Amlodipine	34	Benzonatate	73
Ammonium Chloride	35	Benzoyl Peroxide	73
Amobarbital	35	Benzphetamine	74
Amodiaquine	36	Benztropine	74
Amonafide	36	Benzydamine	75
Amoxapine	37	Bepotastine	75
Amoxicillin	37	Beractant	76
Amphetamine	38	Besifloxacin	76
Amphotericin B	39	Besilesomab	76
		Beta-Carotene	77
		Bethanechol	77
		Betahistine	77
		Betaine	77
		Betamethasone	78
		Betaxolol	79
		Bethanechol	80
		Bevacizumab	81
		Bexarotene	81
		Bezafibrate	82
		Bicalutamide	83
		Bimatoprost	83
		Biotin (Vitamin B ₇)	84
		Biperiden	84
		Bisacodyl	85
		Bismuth	86
		Bisoprolol	86
		Bivalirudin	87
		Bleomycin	88
		Bortezomib	89
		Bosentan	89
		Botulinum Toxin Type A	90
		Botulinum Toxin Type B	91
		Botulism Immune Globulin (Intravenous-Human)	91
		Brimonidine	92
		Brinzolamide	92
		Bromazepam	93
		Bromfenac	94
		Bromocriptine	94
		Bromperidol	95
		Brompheniramine	95
		Brotizolam	96
		Budesonide	96
		Bumetanide	98
		Bupivacaine	98
		Buprenorphine	99
		Bupropion	100
		Buserelin	101
		Buspirone	102
		Busulfan	102
		Butabarbital	103
		Butenafine	104
		Butoconazole	104
		Butorphanol	104
		C	
		Cabazitaxel	106
		Cabergoline	106
		Caffeine	107
		Calcipotriene	108
		Calcitonin	108
		Calcitriol (1 α ,25-dihydroxyvitamin D ₃)	109
		Calcium Acetate	109
		Calcium Carbonate	110
		Calcium Chloride	111
		Calcium Citrate	111
		Calcium Glubionate	111
		Calcium Gluconate	112
		Calcium Lactate	112
		Calcium Phosphate, Tribasic	113
		Calfactant	113
		Canakinumab	113
		Candesartan	114
		Cantharidin	115
		Capecitabine	115
		Capreomycin	116
		Capsaicin	116
		Captopril	116
		Carbachol	117
		Carbamazepine	118
		Carbamide Peroxide	119
		Carbenicillin	119
		Carbidopa	120
		Carbidopa and Levodopa	120
		Carbinoxamine	121
		Carboplatin	121
		Carboprost Tromethamine	122
		Carboxymethylcellulose (Carmellose)	123
		Carglumic Acid	123
		Carisoprodol	123
		Carmustine	124
		Carteolol	124
		Carvedilol	125
		Caspofungin	126
		Catumaxomab	127
		Cefaclor	127
		Cefadroxil	128
		Cefazolin	129
		Cefdinir	129
		Cefditoren	130

Cefepime	130	Cyanocobalamin (Vitamin B ₁₂)	188	Dornase Alpha	242
Cefixime	131	Cyclizine	189	Dorzolamide	242
Cefotaxime	132	Cyclobenzaprine	189	Doxacurium	243
Cefotetan	132	Cyclopentolate	190	Doxapram	244
Cefoxitin	133	Cyclophosphamide	190	Doxazosin	244
Cefpodoxime	133	Cycloserine	191	Doxepin	245
Cefprozil	134	Cyclosporine	191	Doxercalciferol	246
Ceftaroline Fosamil	134	Cyproheptadine	192	Doxorubicin	246
Ceftazidime	135	Cyproterone	193	Doxycycline	248
Ceftibuten	136	Cysteamine (Mercaptamine)	194	Doxylamine	248
Ceftizoxime	136	Cysteine	194	Dronabinol	249
Ceftriaxone	137	Cytarabine	194	Dronedarone	249
Cefuroxime	138	Cytomegalovirus Immune Globulin (Intravenous-Human)	195	Droperidol	250
Celecoxib	139			Drotrecogin Alpha	251
Cephalexin	140	Dabigatran Etxilate	196	Duloxetine	251
Cephradine	140	Dacarbazine	196	Dutasteride	252
Cerebrolysin	141	Daclizumab	197	Dyclonine	253
Certolizumab Pegol	141	Dactinomycin	197	Dyphylline	253
Cetirizine	142	Dalfampridine (4-Aminopyridine)	198		
Cetuximab	142	Dalteparin	198	Ebastine	254
Cetylpyridinium	143	Danaparoid	199	Ecallantide	254
Cevimeline	143	Danazol	200	Echinacea (<i>Echinacea purpurea</i> (L) Moench)	255
Chamomile (<i>Matricaria chamomilla</i> L)	144	Dantrolene	200	Echthiophate Iodide	255
Charcoal, activated	144	Dapsone	201	Econazole	256
Chenodiol (Chenodeoxycholic Acid)	145	Daptomycin	201	Eculizumab	256
Chloral Hydrate	145	Darbepoetin Alpha	202	Edetate Calcium Disodium	257
Chlorambucil	146	Darifenacin	202	Edrophonium	257
Chloramphenicol	146	Darunavir	203	Efalizumab	258
Chlordiazepoxide	147	Dasatinib	204	Eflavirenz	258
Chlorhexidine Gluconate	148	Daunorubicin Hydrochloride	204	Eflornithine	259
Chlorprocaine	149	Decitabine	205	Eletriptan	259
Chloroquine	149	Deferasirox	205	Eltrombopag	260
Chlorothiazide	150	Deferoxamine	206	Emtricitabine	261
Chlorpheniramine	151	Degarelix	207	Enalapril	261
Chlorpromazine	151	Delavirdine	207	Enfuvirtide	262
Chlorpropamide	152	Demeclocycline	208	Enoxaparin	263
Chlorthalidone	153	Denileukin Diftitox	208	Entacapone	263
Chlorzoxazone	154	Denosumab	209	Entecavir	264
Cholecalciferol (Vitamin D ₃)	154	Desflurane	209	Ephedrine	265
Cholestyramine Resin	155	Desipramine	209	Epinastine	265
Choline Magnesium Trisalicylate	155	Desloratadine	210	Epinephrine (Adrenaline)	266
Choriogonadotropin Alpha	156	Desmopressin	211	Epirubicin	267
Chorionic Gonadotropin (Human)	156	Desonide	212	Eplerenone	267
Ciclesonide	157	Desoximetastone	212	Epoetin Alpha	268
Ciclopirox	157	Desvenlafaxine	213	Epoetin Theta	269
Cidofovir	158	Dexamethasone	213	Epoprostenol	269
Cilazapril	158	Dexchlorpheniramine	215	Eprosartan	270
Cilostazol	159	Dexlansoprazole	215	Epitibatide	270
Cimetidine	160	Dexmedetomidine	216	Eptotermin Alpha	271
Cinacalcet	161	Dexmethyphenidate	216	Ergocalciferol (Vitamin D ₂)	271
Cinnarizine	161	Dexpantenol	217	Ergoloid Mesylates	272
Ciprofloxacin	162	Dextrazoxane	217	Ergonovine (Ergometrine)	273
Cisapride	163	Dextran	218	Ergotamine	273
Cisatracurium	164	Dextroamphetamine	218	Eribulin Mesylate	274
Cisplatin	164	Dextromethorphan	219	Erlotinib	274
Citalopram	165	Dextrose (D-Glucose)	219	Ertapenem	275
Citicoline	166	Diazepam	220	Erythromycin	276
Cladribine	167	Diazoxide	221	Escitalopram	277
Clarithromycin	167	Dibucaine (Cinchocaine)	221	Eslicarbazepine	278
Clemastine	168	Dichlorphenamide	222	Esmolol	279
Clindamycin	169	Diclofenac	222	Esomeprazole	279
Clobazam	170	Dicloxacillin	223	Estazolam	280
Clobetasol	170	Dicyclomine	224	Estradiol	281
Clocortolone	171	Didanosine	224	Estramustine	282
Clodronate	171	Diethylpropion	225	Estrogens (Conjugated)	282
Clofarabine	172	Difforasone	225	Eszopiclone	283
Clomiphene (Clomifene)	172	Diffunisal	226	Etanercept	284
Clomipramine	173	Diffuprednate	227	Ethacrynic Acid	285
Clonazepam	174	Digoxin	227	Ethambutol	285
Clonidine	175	Dihydroergotamine	228	Ethionamide	286
Clopidogrel	175	Dihydrorotachysterol	228	Ethosuximide	286
Clorazepate	176	Diloxanide Furoate	229	Ethotoin	287
Clotrimazole	177	Diltiazem	229	Ethyl Chloride	287
Cloxacillin	178	Dimenhydrinate	230	Etidronate Disodium	287
Clozapine	178	Dimercaprol	230	Etodolac	288
Coal Tar	179	Dimethyl Sulfoxide	231	Etomidate	289
Cocaine	179	Dinoprostone	231	Etonogestrel	289
Codeine	180	Diphenhydramine	232	Etoposide	290
Colchicine	181	Dipivefrin	233	Etoricoxib	290
Colesevelam	182	Dipyridamole	233	Etravirine	291
Colestipol	182	Dipyrene (Metamizole)	234	Everolimus	292
Colistimethate	183	Disopyramide	235	Exemestane	293
Collagen Hemostat	183	Disulfiram	235	Exenatide	293
Collagenase	183	Dobutamine	236	Ezetimibe	294
Conestat Alpha	184	Docetaxel	236		
Conivaptan	184	Docosanol	238	F	
Corifoliotropin Alpha	185	Docusate	238	Factor IX Complex (Human)	295
Corticoirelin	185	Dofetilide	238	Factor VIIa (Recombinant)	295
Corticotropin	185	Dolasetron	239	Famciclovir	296
Cortisone	186	Domperidone	239	Famotidine	296
Cosyntropin (Tetracosactide)	187	Donepezil	240	Febuxostat	297
Cromolyn	187	Dopamine	241	Felbamate	297
Crotamiton	188	Doripenem	241	Felodipine	298
				Fenofibrate	299

Fenoldopam	300	Heparin	356	L	
Fenoprofen	300	Hepatitis A Vaccine	357	L-Lysine	411
Fentanyl	301	Hepatitis B Immune Globulin	357	Labetalol	411
Ferric Gluconate	302	Hepatitis B Vaccine	358	Lacidipine	412
Ferrous Fumarate	302	Histrelin	358	Lactase	412
Ferrous Sulfate	303	Homatropine	359	Lactobacillus	412
Ferumoxytol	303	Hops (<i>Humulus lupulus</i> L.)	359	Lactulose	413
Fesoterodine	304	Human papillomavirus quadrivalent (types 6, 11, 16, and 18) vaccine, recombinant	359	Lamivudine	413
Fexofenadine	304	Huperzine A (<i>Huperzia serrata</i> Thunb)	360	Lamotrigine	414
Filgrastim	305	Hyaluronidase (Human Recombinant)	360	Lanreotide	414
Finasteride	305	Hydralazine	361	Lansoprazole	415
Fingolimod	306	Hydrochlorothiazide	361	Lanthanum	416
Flavocoxid	307	Hydrocodone	362	Lapatinib	416
Flavoxate	307	Hydrocortisone	363	Laroidase	417
Flecainide	308	Hydromorphone	364	Lasofloxiene	417
Floxuridine	308	Hydroquinone	365	Latanoprost	418
Flucloxacillin (Flxacillin)	309	Hydroxycobalamin (Vitamin B _{12a})	365	Leftunomide	419
Fluconazole	309	Hydroxychloroquine	366	Lenalidomide	419
Flucytosine	310	Hydroxypropyl Cellulose	367	Lepirudin	420
Fludarabine	311	Hydroxypropyl Methylcellulose (Hypromellose)	367	Letrozole	420
Fludrocortisone	311	Hydroxyurea (Hydroxycarbamide)	367	Leucovorin (Folinic Acid)	421
Flumazenil	312	Hydroxyzine	368	Leuprolide	422
Flunarizine	313	Hyosciamine	368	Levalbuterol	422
Flunisolide	313			Levetiracetam	423
Fluocinolone	314	I		Levobunolol	423
Fluocinonide	314	Ibandronate	370	Levobupivacaine	424
Fluoride	315	Ibritumomab	370	Levocabastine	424
Fluorometholone	315	Ibuprofen	371	Levocarnitine	425
Fluorouracil	316	Ibutilide	372	Levocetirizine	425
Fluoxetine	317	Idarubicin	372	Levofloxacin	426
Fluoxymesterone	318	Idebenone	373	Levonorgestrel	426
Flupenthixol	318	Ifosfamide	374	Levorphanol	427
Fluphenazine	319	Iloperidone	374	Levothyroxine	428
Flurazepam	320	Iloprost	375	Lidocaine	428
Flurbiprofen	320	Imatinib	375	Lincomycin	430
Flutamide	321	Imiglucerase	376	Linezolid	430
Fluticasone	322	Imipenem and Cilastatin	377	Liothyronine	431
Fluvastatin	323	Imipramine	378	Liotrix	431
Fluvoxamine	324	Imiquimod	378	Liraglutide	432
Folic Acid (Vitamin B ₉)	325	Immune Globulin, Intramuscular	379	Lisdexamfetamine	432
Follitropin Alpha	325	Immune Globulin, Intravenous	379	Lisinopril	433
Follitropin Beta	326	Inamrinone	380	Lisuride	434
Fomepizole	326	Indacaterol	381	Lithium	434
Fondaparinux	327	Indapamide	381	Lodoxamide	435
Formoterol	327	Indinavir	382	Lomefloxacin	435
Fosamprenavir	328	Indomethacin	383	Lomustine	436
Fosaprepitant	329	Infliximab	384	Loperamide	436
Foscarnet	330	Influenza Virus Vaccine	385	Loprazolam	437
Fosfomycin	330	Insulin	385	Loracarbef	437
Fosinopril	331	Interferon Alpha-2a	386	Loratadine	438
Fosphenytoin	332	Interferon Alpha-2b	387	Lorazepam	439
Frovatriptan	332	Interferon Alpha-n3	388	Lormetazepam	439
Furosemide	333	Interferon Alphacon-1	388	Lornoxicam	440
		Interferon Beta-1a	388	Losartan	440
G		Interferon Beta-1b	389	Loteprednol	441
Gabapentin	334	Interferon Gamma-1b	390	Lovastatin	442
Galantamine	334	Iodipamide Meglumine	390	Loxapine	442
Gallium Nitrate	335	Iodixanol	390	Lubiprostone	443
Galsulfase	336	Iodoquinol	391	Lurasidone	443
Ganciclovir	336	Iohexol	391	Lutropin Alpha	444
Ganirelix	337	Iopamidol	391		
Gasfloxacin	337	Iothalamate Meglumine	392	M	
Gefitinib	337	Ioversol	392	Mafenide	445
Gemcitabine	338	Ioxaglate Meglumine and Ioxaglate Sodium	392	Magnesium Chloride	445
Gemfibrozil	339	Ipecac Syrup	393	Magnesium Citrate	445
Gemifloxacin	339	Ipratropium	393	Magnesium Glucoheptonate	446
Gemtuzumab Ozogamicin	340	Irbesartan	394	Magnesium Gluconate	446
Gentamicin	341	Irinotecan	394	Magnesium Hydroxide	446
Ginkgo biloba (<i>Ginkgo biloba</i> L.)	342	Iron Dextran Complex	395	Magnesium L-Aspartate Hydrochloride	447
Ginseng (<i>Panax ginseng</i> CA Meyer)	342	Iron Sucrose	396	Magnesium L-lactate	447
Glatiramer Acetate	343	Isocarboxazid	396	Magnesium Oxide	448
Glclazide	344	Isoflurane	397	Magnesium Salicylate	448
Glimepiride	344	Isoniazid	397	Magnesium Sulfate	448
Glipizide	345	Isoproterenol	398	Malathion	449
Glucagon	346	Isosorbide Dinitrate	398	Maltodextrin	449
Glyburide (Glibenclamide)	346	Isosorbide Mononitrate	399	Manganese	449
Glycopyrrolate	347	Isotretinoin	400	Mannitol	450
Gold Sodium Thiomalate	347	Isoxsuprine	401	Maprotiline	450
Golimumab	348	Isradipine	401	Maraviroc	451
Gonadorelin	348	Itraconazole	402	Measles Virus Vaccine (Live)	451
Goserelin	349	Ivabradine	403	Measles, Mumps, and Rubella Vaccines (Combined)	452
Granisetron	349	Ivermectin	403	Measles, Mumps, Rubella and Varicella Virus Vaccine	452
Griseofulvin	350	Ixabepilone	404	Mebendazole	453
Guafenesin	351			Mecamylamine	453
Guanabenz	351	J		Mecasermin	454
Guanfacine	352	Japanese Encephalitis Virus Vaccine (Inactivated)	405	Mechlorethamine	454
		K		Meclizine (Meclizine)	454
<i>Haemophilus</i> b Conjugate Vaccine	353	Kanamycin	406	Meclofenamate	455
Halcinonide	353	Ketamine	406	Medium Chain Triglycerides	456
Halobetasol	353	Ketoconazole	407	Medroxyprogesterone	456
Haloperidol	354	Ketoprofen	408	Mefenamic Acid	457
Halothane	355	Ketorolac	409	Mefloquine	457
Hawthorn (<i>Crataegus oxyacantha</i> L.)	355	Ketotifen	410		
Hemin	356				

Megestrol	458	Nalmefene	509	Papaverine	565
Meloxicam	459	Naloxone	510	Paregoric	565
Melphalan	460	Naltrexone	510	Paricalcitol	566
Memantine	460	Nandrolone	511	Paromomycin	566
Meningococcal Group C-CRM197 Conjugate Vaccine	461	Naphazoline	511	Paroxetine	567
Meningococcal Polysaccharide (Groups A/C/Y and W-135)		Naproxen	512	Pazopanib	568
Diphtheria Toxoid Conjugate Vaccine	461	Naratriptan	513	Pegademase Bovine	568
Meningococcal Polysaccharide Vaccine (Groups A/C/Y and W-135)	461	Natalizumab	513	Pegaptanib	569
Menotropins	462	Natamycin	514	Pegaspargase	569
Mepenzolate	462	Nateglinide	514	Pegfilgrastim	570
Meperidine (Pethidine)	463	Nebivolol	515	Peginterferon Alpha-2a	570
Mephobarbital (Methylphenobarbital)	463	Nedocromil	516	Peginterferon Alpha-2b	571
Mepivacaine	464	Nefazodone	516	Pegloticase	572
Meprobamate	464	Nelarabine	517	Pegvisomant	572
Mercaptopurine	465	Nelfinavir	518	Pemetrexed	573
Meropenem	465	Neomycin	518	Penciclovir	573
Mesalamine (Mesalazine)	466	Neostigmine	519	Penicillamine	573
Mesna	467	Nepafenac	520	Penicillin G Benzathine	574
Metaprotenerol (Orciprenaline)	467	Nesiritide	520	Penicillin G Procaine	574
Metaxalone	468	Nevirapine	521	Penicillin G (Parenteral/Aqueous)	575
Metformin	468	Niacin (Nicotinic Acid; Vitamin B ₃)	521	Penicillin V Potassium	575
Methacholine	469	Niacinamide (Nicotinamide)	522	Pentamidine	576
Methadone	469	Nicardipine	522	Pentastarch	577
Methamphetamine	470	Nicergoline	523	Pentazocine	577
Methazolamide	470	Nicotine	524	Pentobarbital	577
Methenamine	471	Nifedipine	524	Pentosan Polysulfate Sodium	578
Methimazole (Thiamazole)	471	Nilotinib	525	Pentostatin	578
Methocarbamol	472	Nilutamide	526	Pentoxifylline	579
Methohexital	472	Nimesulide	527	Pergolide	579
Methotrexate	473	Nimodipine	528	Periciazine (Propiciazine)	580
Methotrimeprazine (Levomepromazin)	474	Nisoldipine	528	Perindopril Erbumine	581
Methoxsalen	474	Nitazoxanide	529	Permethrin	581
Methscopolamine	475	Nitisinone	529	Perphenazine	582
Methsuximide	475	Nitrazepam	530	Phenazopyridine	583
Methyclothiazide	476	Nitrendipine	530	Phendimetrazine	583
Methylcellulose	476	Nitric Oxide	531	Phenelzine	583
Methylidopa	476	Nitrofurantoin	531	Phenindamine	584
Methylene Blue	477	Nitrofurazone (Nitrofuraz)	532	Phenobarbital	584
Methylethylgonovine	477	Nitroglycerin (Glyceryl Trinitrate)	532	Phenol	585
Methylphenidate	478	Nitroprusside	533	Phenoxybenzamine	585
Methylprednisolone	479	Nizatidine	534	Phentermine	586
Methyltestosterone	479	Nonoxonyl 9	534	Phentolamine	586
Methysergide	480	Norepinephrine (Noradrenaline)	534	Phenylephrine	587
Metipranolol	480	Norethindrone (Norethisterone)	535	Phenytoin	588
Metoclopramide	481	Norfloxacin	535	Physostigmine	589
Metolazone	481	Nortriptyline	536	Phytonadione (Vitamin K ₁)	589
Metoprolol	482	Nystatin	537	Pilocarpine	589
Metronidazole	483	O		Pimecrolimus	590
Metirapone	484	Octreotide	539	Pimozide	591
Metirosine	484	Ofatumumab	540	Pinaverium	591
Mexiletine	484	Ofloxacin	540	Pindolol	592
Mianserin	485	Olanzapine	541	Pioglitazone	593
Micafungin	485	Olmesartan	542	Pipecuronium	593
Miconazole	486	Olopatadine	543	Piperacillin	594
Midazolam	487	Olsalazine	543	Piperazine	594
Midodrine	488	Omalizumab	544	Pipotiazine	595
Mifamuride	488	Omega-3-Acid Ethyl Esters	544	Piracetam	595
Mifepristone	489	Omeprazole	545	Pirbuterol	596
Miglitol	489	Ondansetron	546	Piroxicam	596
Miglustat	490	Oprelvekin	546	Pitavastatin	597
Milnacipran	490	Orlistat	547	Pivampicillin	598
Milrinone	491	Orphenadrine	547	Pizotifen (Pizotyline)	598
Minocycline	491	Oseltamivir	548	Plasma Protein Fraction	599
Minoxidil	492	Oxacillin	548	Plerixafor	599
Mirtazapine	493	Oxaliplatin	549	Pneumococcal Conjugate Vaccine (7-Valent)	600
Misoprostol	494	Oxandrolone	550	Podofilox	600
Mitomycin	494	Oxaprozin	550	Podophyllum Resin	600
Mitotane	495	Oxatomide	551	Poliovirus Vaccine (Inactivated)	601
Mitoxantrone	495	Oxazepam	552	Poly-L-Lactic Acid	601
Mivacurium	496	Oxcarbazepine	552	Polycarbophil	601
Moclobemide	497	Oxiconazole	553	Polyethylene Glycol 3350	601
Modafinil	497	Oxprenolol	553	Polymyxin B	602
Moexipril	498	Oxybutynin	554	Polysaccharide-Iron Complex	602
Molindone	499	Oxycodone	555	Poractant Alpha	602
Mometasone	499	Oxygen	556	Porfimer	603
Monobenzone	500	Oxymetazoline	556	Posaconazole	603
Montelukast	500	Oxymetholone	556	Potassium Acetate	604
Moricizine	501	Oxymorphone	557	Potassium Acid Phosphate	604
Morphine Sulfate	501	Oxytetracycline	557	Potassium Bicarbonate	605
Morrhuate Sodium	502	Oxytocin	558	Potassium Chloride	605
Moxifloxacin	503	P		Potassium Citrate	606
Mupirocin	503	Paclitaxel	559	Potassium Gluconate	606
Muromonab-CD3	504	Palifermin	560	Potassium Iodide	606
Mycophenolate	504	Paliperidone	560	Potassium p-Aminobenzoate	607
N		Palivizumab	561	Potassium Phosphate	607
Nabilone	506	Palonosetron	561	Povidone-Iodine	607
Nabumetone	506	Pamidronate	561	Pralatrexate	608
Nadolol	507	Pancreatin	562	Pralidoxime	608
Nafarelin	508	Pancrelpase	562	Pramipexole	609
Nafcillin	508	Pancuronium	563	Pramlintide	610
Naftifine	508	Panitumumab	563	Pramoxine	610
Nalbuphine	509	Pantoprazole	564	Prasugrel	610
Nalidixic Acid	509	Pantothenic Acid (Vitamin B ₅)	565	Pravastatin	611

World Guide for Drug Use and Pharmacogenomics

Drugs

A

Abacavir

Brand Names
Europe

Austria: Ziagen; **Belgium:** Ziagen; **Bulgaria:** Ziagen; **Cyprus:** Ziagen; **Czech Republic:** Ziagen; **Denmark:** Ziagen; **Estonia:** Ziagen; **Finland:** Ziagen; **France:** Ziagen; **Germany:** Ziagen; **Greece:** Ziagen; **Hungary:** Ziagen; **Ireland:** Ziagen; **Italy:** Ziagen; **Latvia:** Ziagen; **Lithuania:** Ziagen; **Luxembourg:** Ziagen; **Malta:** Ziagen; **Netherlands:** Ziagen; **Poland:** Ziagen; **Portugal:** Ziagen; **Romania:** Ziagen; **Slovakia:** Ziagen; **Slovenia:** Ziagen; **Spain:** Ziagen; **Sweden:** Ziagen; **UK:** Ziagen.

North America

Canada: Ziagen; **USA:** Ziagen.

Latin America

Argentina: Abacavir, Filabac, Finecil, Panka, Plusabcir, Zepiril, Ziagenavir; **Brazil:** Ziagenavir; **Mexico:** Ziagenavir.

Asia

Japan: Ziagen.

Drug Combinations

Abacavir and Lamivudine
Abacavir, Lamivudine, and Zidovudine

Chemistry

Abacavir Sulfate: (C₁₄H₁₈N₄O)₂ H₂SO₄. Mw: 670.74. (1S,4R)-4-[2-Amino-6-(cyclopropylamino)-9H-purin-9-yl]-2-cyclopentene-1-methanol sulfate (2:1). CAS-188062-50-2 (1997).

Abacavir Succinate: C₁₄H₁₈N₄O C₄H₄O₄. Mw: 404.42. 2-Cyclopentene-1-methanol, 4-[2-amino-6-(cyclopropylamino)-9H-purin-9-yl]-, (1S-cis)-, butanedioate (1:1). CAS-168146-84-7; CAS-136470-78-5 (abacavir)(1996).



Pharmacologic Category

Anti-Infective Agents; Antiretrovirals; Nucleoside and Nucleotide Reverse Transcriptase Inhibitors. (ATC-Code: J05AF06).

Mechanism of Action

Abacavir is phosphorylated to carbovir triphosphate, which interferes with HIV viral RNA-dependent DNA polymerase, inhibiting viral replication.

Therapeutic Use

Treatment of HIV infections in combination with other antiretroviral agents.

Pregnancy and Lactation Implications

Risk of lactic acidosis and liver damage in pregnant women. Contraindicated during lactation.

Contraindications

Hypersensitivity to abacavir (or carbovir) or any component of the formulation. Hepatic impairment.

Warnings and Precautions

Hypersensitivity reactions have occurred (increased risk in patients with *HLA-B*5701* allele). Lactic acidosis and severe hepatomegaly with steatosis have been reported (caution in risk factors for liver disease). Caution in hepatic dysfunction (contraindicated in moderate-to-severe dysfunction) or in risk factors for coronary heart disease. Immune reconstitution syndrome may develop. May cause redistribution of fat. Should always be used as part of a multidrug regimen.

Adverse Reactions

Cardiovascular: Myocardial infarction.

Central Nervous System: Abnormal dreams, anxiety, depression, dizziness, fatigue, headache, insomnia, lethargy, malaise, paresthesia.

Dermatologic: Erythema multiforme, rash, Stevens-Johnson syndrome, toxic epidermal necrolysis.

Endocrine and Metabolic: Fat redistribution, increased triglycerides, lactic acidosis.

Gastrointestinal: Diarrhea, increased amylase, nausea, pancreatitis, vomiting.

Hematologic: Thrombocytopenia.

Hepatic: Hepatic steatosis, hepatomegaly, hepatotoxicity, increased AST, increased GGT.

Neuromuscular and Skeletal: Musculoskeletal pain.

Miscellaneous: Chills, fever, hypersensitivity reactions, infection.

Pharmacogenetics

Caution and personalized dose adjustment in patients with the following genotypes:

HCP5: rs2395029

HLA-B: *HLA-B*5701*, *HLA-B*1502*, *HLA-B*5801*, *HLA-B18* subtype, *HLA-B39* subtype,

HLA-B44 subtype

HSPA1L: Thr493Met

Other genes that may be involved:

ABCB1; *ABCC6*; *ADH5*; *CRP*; *TERT*; *UGT1A1*; *UGT2B7*

Inhibits:

ABCC1; *ABCC2*

Induces:

CYP2B6

Drug Interactions

Ganciclovir-valganciclovir may enhance the adverse/toxic effect of reverse transcriptase inhibitors (nucleoside)(hematologic toxicity). Protease inhibitors may decrease the serum concentration of abacavir. Ribavirin may enhance the hepatotoxic effect of reverse transcriptase inhibitors (nucleoside)(lactic acidosis).

Nutrition/Nutraceutical Interactions

Ethanol: May increase risk of toxicity.

Dosage

Dosage Forms: Solution, oral: 20 mg/mL (240 mL)(strawberry-banana flavor). Tablet: 300 mg. [Oral administration].

Pharmacokinetics and Pharmacodynamics

Absorption: Rapid and extensive.

Distribution: V_d: 0.86 L/kg.

Protein binding: 50%.

Metabolism: Via alcohol dehydrogenase and glucuronosyltransferase to inactive carboxylate and glucuronide metabolites.

Bioavailability: 83%.

Half-life: 1.5 hours.

Time to peak: 0.7-1.7 hours.

Excretion: Urine (primarily), feces.

Special Considerations

Diet: May be taken with or without food.

Monitoring Parameters: CBC, serum creatine kinase, CD4 count, HIV RNA plasma levels, serum transaminases, triglycerides, serum amylase, *HLA-B*5701* genotype status prior to initiation of therapy, signs and symptoms of hypersensitivity.

Abacavir and Lamivudine

Brand Names

Europe

Austria: Kivexa; **Belgium:** Kivexa; **Bulgaria:** Kivexa; **Cyprus:** Kivexa; **Czech Republic:** Kivexa; **Denmark:** Kivexa; **Estonia:** Kivexa; **Finland:** Kivexa; **France:** Kivexa; **Germany:** Kivexa; **Greece:** Kivexa; **Hungary:** Kivexa; **Ireland:** Kivexa; **Italy:** Kivexa; **Latvia:** Kivexa; **Lithuania:** Kivexa; **Luxembourg:** Kivexa; **Malta:** Kivexa; **Netherlands:** Kivexa; **Poland:** Kivexa; **Portugal:** Kivexa; **Romania:** Kivexa; **Slovakia:** Kivexa; **Slovenia:** Kivexa; **Spain:** Kivexa; **Sweden:** Kivexa; **UK:** Kivexa.

North America

Canada: Kivexa; **USA:** Epzicom.

Latin America

Argentina: Kivexa; **Brazil:** Kivexa; **Mexico:** Kivexa.

Asia

Japan: Epzicom.

Drug Combinations

Abacavir, Lamivudine, and Zidovudine

Chemistry

Abacavir Sulfate: (C₁₄H₁₈N₄O)₂ H₂SO₄. Mw: 670.74. (1S,4R)-4-[2-Amino-6-(cyclopropylamino)-9H-purin-9-yl]-2-cyclopentene-1-methanol sulfate (2:1). CAS-188062-50-2 (1997).

Lamivudine: C₈H₁₀N₂O₃S. Mw: 229.26. (1) 2(1H)-Pyrimidinone, 4-amino-1-[2-(hydroxymethyl)-1,3-oxathiolan-5-yl]-, (2R-cis)-; (2)(-)-1-[(2R,5S)-2-(Hydroxymethyl)-1,3-oxathiolan-5-yl]cytosine. CAS-134678-17-4 (1992).



Pharmacologic Category

Anti-infective Agents, Antiretrovirals, Nucleoside and Nucleotide Reverse Transcriptase Inhibitors. (ATC-Code: J05AR02).

Mechanism of Action

Abacavir phosphorylated to carbovir triphosphate interferes with HIV viral RNA-dependent DNA polymerase, inhibiting viral replication. Triphosphorylated lamivudine inhibits HIV reverse transcription via viral DNA chain termination. Inhibits RNA-dependent DNA polymerase activities of reverse transcriptase.

Therapeutic Use

Treatment of HIV infections in combination with other antiretroviral agents.

Contraindications

Hypersensitivity to abacavir, lamivudine, or any component of the formulation. Renal (CrCl <50 mL/minute) and hepatic impairment.

Warnings and Precautions

May cause redistribution of fat. Hypersensitivity reactions have occurred (increased risk in patients with *HLA-B*5701* allele). Immune reconstitution syndrome might be developed. Lactic acidosis and hepatomegaly with steatosis have been reported. Clinical exacerbations after discontinuing therapy for chronic hepatitis B in HIV patients have occurred. Caution in combination with interferon alfa with or without ribavirin in HIV/HBV-coinfected patients. Caution in renal dysfunction (contraindicated in CrCl <50 mL/minute or in risk factors for coronary heart disease). Avoid use in children.

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Adverse Reactions

Central Nervous System: Paresthesia, peripheral neuropathy, seizure.
Dermatologic: Alopecia, erythema multiforme, urticaria.
Endocrine and Metabolic: Fat redistribution, hyperglycemia, lactic acidosis, Stevens-Johnson syndrome.
Gastrointestinal: Pancreatitis, stomatitis.
Hematologic: Anemia, aplastic anemia.
Hepatic: Hepatic steatosis, hepatitis B exacerbation.
Neuromuscular and Skeletal: Increased CPK, muscle weakness, rhabdomyolysis, weakness.
Respiratory: Abnormal breath sounds, wheezing.
Miscellaneous: Anaphylaxis, hypersensitivity reaction, lymphadenopathy, splenomegaly.

Pharmacogenetics

Caution and personalized dose adjustment in patients with the following genotypes:
HCP5: rs2395029
HFE: C187G
HLA-B: HLA-B*5701, HLA-B*1502, HLA-B*5801, HLA-B18 subtype, HLA-B39 subtype, HLA-B44 subtype
HSPA1L: Thr493Met
Other genes that may be involved:
ABCB1; ABCC6; ABCG2; ADHs; CRP; ICAM1; UGT1A1; UGT2B7
Inhibits:
ABCC1; ABCC2
Induces:
CYP2B6

Drug Interactions

Ganciclovir-valganciclovir may enhance the adverse/toxic effect of reverse transcriptase inhibitors (nucleoside)(hematologic toxicity). Protease inhibitors may decrease the serum concentration of abacavir. Ribavirin may enhance the hepatotoxic effect of reverse transcriptase inhibitors (nucleoside)(lactic acidosis). Trimethoprim may decrease the excretion of lamivudine. Lamivudine may diminish the therapeutic effect of zalcitabine.

Dosage

Dosage Forms: Tablet: Abacavir 600 mg and lamivudine 300 mg. [Oral administration].
Dose adjustment in special conditions: In renal (use not recommended) or hepatic (use contraindicated) impairment.

Abacavir, Lamivudine, and Zidovudine

Brand Names

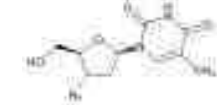
Europe
Austria: Trizivir; Belgium: Trizivir; Bulgaria: Trizivir; Cyprus: Trizivir; Czech Republic: Trizivir; Denmark: Trizivir; Estonia: Trizivir; Finland: Trizivir; France: Trizivir; Germany: Trizivir; Greece: Trizivir; Hungary: Trizivir; Ireland: Trizivir; Italy: Trizivir; Latvia: Trizivir; Lithuania: Trizivir; Luxembourg: Trizivir; Malta: Trizivir; Netherlands: Trizivir; Poland: Trizivir; Portugal: Trizivir; Romania: Trizivir; Slovakia: Trizivir; Slovenia: Trizivir; Spain: Trizivir; Sweden: Trizivir; UK: Trizivir.
North America
Canada: Trizivir; USA: Trizivir.
Latin America
Argentina: Tricivir, Trividin, Zidomuv; Mexico: Trizivir.

Chemistry

Abacavir Sulfate: (C₈H₁₀N₄O)₂·H₂SO₄. Mw: 670.74. (1S,4R)-4-[2-Amino-6-(cyclopropylamino)-9H-purin-9-yl]-2-cyclopentene-1-methanol sulfate (2:1). CAS-188062-50-2 (1997).
Lamivudine: C₈H₁₀N₄O₅S. Mw: 229.26. (1) 2(1H)-Pyrimidinone, 4-amino-1-[2-(hydroxymethyl)-1,3-oxathiolan-5-yl]-, (2R-cis)-; (2)-1-1-[[2R,5S)-2-(Hydroxymethyl)-1,3-oxathiolan-5-yl]cytosine. CAS-134678-17-4 (1992).



Zidovudine: C₈H₁₀N₄O₅. Mw: 267.24. (1) Thymidine, 3'-azido-3'-deoxy-; (2) 3'-Azido-3'-deoxythymidine. CAS-30516-87-1 (1987).



Pharmacologic Category

Anti-infective Agents, Antiretrovirals, Nucleoside and Nucleotide Reverse Transcriptase Inhibitors. (ATC-Code: J05AR04).

Mechanism of Action

Inhibition of reverse transcriptase via DNA chain termination after incorporation of the nucleoside analog as well as delay of emerging mutations conferring resistance.

Therapeutic Use

Treatment of HIV infections in combination with other antiretroviral agents.

Contraindications

Hypersensitivity to abacavir, lamivudine, zidovudine, or any component of the formulation. Renal (CrCl ≤50 mL/minute) or hepatic impairment.

Warnings and Precautions

May cause redistribution of fat. Hematologic toxicity may occur (neutropenia, anemia; use with caution in bone marrow compromise). Hypersensitivity reactions have occurred (increased risk in patients with HLA-B*5701 allele). Immune reconstitution syndrome may develop. Lactic acidosis and severe hepatomegaly with steatosis have been reported (caution in risk factors for liver disease). Clinical exacerbations after discontinuing therapy for chronic hepatitis B in HIV patients have occurred. Caution in risk factors for coronary heart disease. Risk of myopathy and myositis (with prolonged use). Should always be used as part of a multidrug regimen. Should not be used in CrCl ≤50 mL/minute. Caution in combination with interferon alfa with or without ribavirin in HIV/HBV coinfectd patients. Avoid use in children.

Adverse Reactions

Central Nervous System: Anxiety, depression, fatigue, headache, malaise.
Dermatologic: Rash.
Endocrine and Metabolic: Fat redistribution, increased triglycerides.
Gastrointestinal: Diarrhea, increased amylase, nausea, pancreatitis, vomiting.
Hematologic: Neutropenia.
Hepatic: Increased ALT, increased GGT.

Neuromuscular and Skeletal: Increased CPK.
Miscellaneous: Ear/nose/throat infection, chills, fever, hypersensitivity (based on abacavir component), immune reconstitution syndrome, viral infection.

Pharmacogenetics

Caution and personalized dose adjustment in patients with the following genotypes:
ABCC4: C1612T, G3463A, G3724A, T4131G
ABCC2: C421A and G34A
CYP2A6: CYP2A6*4, CYP2A6*7, CYP2A6*9
CYP2C9: CYP2C9*2, CYP2C9*3, CYP2C9*5
CYP2C19: CYP2C19*2, CYP2C19*3, CYP2C19*4, CYP2C19*5, CYP2C19*6, CYP2C19*7, CYP2C19*8, CYP2C19*17
CYP3A4: CYP3A4*1, CYP3A4*1B, CYP3A4*2, CYP3A4*3, CYP3A4*4, CYP3A4*5, CYP3A4*6, CYP3A4*8, CYP3A4*11, CYP3A4*12, CYP3A4*13, CYP3A4*15, CYP3A4*17, CYP3A4*18, CYP3A4*19
HCP5: rs2395029
HFE: C187G
HLA-B: HLA-B*5701, HLA-B*1502, HLA-B*5801, HLA-B18 subtype, HLA-B39 subtype, HLA-B44 subtype
HSPA1L: Met493Thr
UGT2B7: UGT2B7*2 (His268Tyr)
Other genes that may be involved:
ABCB1; ABCC6; ADHs; ICAM; UGT1A1
Inhibits:
ABCC1; ABCC2
Induces:
CYP2B6

Drug Interactions

Acydovir-valacydovir may enhance the CNS-depressant effect of zidovudine. Doxorubicin and interferons may enhance the adverse/toxic effect of zidovudine. Doxorubicin may diminish the therapeutic effect of zidovudine. Fluconazole and probenecid may decrease the metabolism of zidovudine. Ganciclovir-valganciclovir may enhance the adverse/toxic effect of reverse transcriptase inhibitors (nucleoside). Interferons and valproic acid may decrease the metabolism of zidovudine. Methadone may increase the serum concentration of zidovudine. Protease inhibitors may decrease the serum concentration of zidovudine, and of abacavir. Ribavirin may enhance the hepatotoxic effect of reverse transcriptase inhibitors (nucleoside). Rifamycin derivatives may increase the metabolism of zidovudine (exceptions: rifabutin). Zidovudine may diminish the therapeutic effect of stavudine. Trimethoprim may decrease the excretion of lamivudine. Lamivudine may diminish the therapeutic effect of zalcitabine.

Dosage

Dosage Forms: Tablet: Abacavir 300 mg, lamivudine 150 mg, and zidovudine 300 mg. [Oral administration].
Dose adjustment in special conditions: In renal (CrCl ≤50 mL/minute: avoid use) or hepatic (use contraindicated) impairment.

Abarelix

Brand Names

Europe
Austria: Plenaxis; Germany: Plenaxis; Netherlands: Plenaxis.

Chemistry

Abarelix: C₂₈H₃₆ClN₄O₄. Mw: 1416.06. N-Acetyl-3-(2-naphthyl)-D-alanyl-4-chloro-D-phenylalanyl-3-(3-pyridyl)-D-alanyl-L-seryl-N-methyl-L-tyrosyl-D-asparaginy-L-leucyl-N⁶-isopropyl-L-lysyl-L-prolyl-D-alaninamide. CAS-183552-38-7 (1997).

Pharmacologic Category

Antineoplastic Agents; Endocrine Therapy; Gonadotropin Releasing Hormone Antagonists. (ATC-Code: L02BX01).

Therapeutic Use

Palliative treatment of advanced symptomatic prostate cancer.

Pregnancy and Lactation Implications

Should not be used in women.

Contraindications

Hypersensitivity to abarelix or any ingredient in the formulation. Not indicated in women or pediatric patients.

Warnings and Precautions

Risk of allergic reactions. May cause fetal harm. Increases in serum AST (SGOT) or ALT (SGPT) concentrations. Prolongation of QTc interval. Risk of decreased bone mineral density following chronic therapy with GnRH antagonists and agonists.

Adverse Reactions

Cardiovascular: Hot flushes, peripheral edema.
Central Nervous System: Dizziness, fatigue, headache, sedation, sleep disturbance.
Gastrointestinal: Constipation, diarrhea, nausea.
Neuromuscular and Skeletal: Back pain.
Renal: Dysuria, frequent urination, urinary retention, urinary tract infection.
Respiratory: Upper respiratory tract infection.
Miscellaneous: Breast enlargement, breast pain or nipple tenderness.

Pharmacogenetics

Genes that may be involved:
GNRHR

Drug Interactions

No formal drug interaction studies to date. Metabolism unlikely to involve CYP isoenzymes.

Dosage

Dosage Forms: Injection, powder for reconstitution. [I.M. administration].

Pharmacokinetics and Pharmacodynamics

Absorption: Slow.
Distribution: Extensive (approximately 4040 L).
Protein binding: 96-99%.
Metabolism: Unlikely to involve CYP isoenzymes.
Half-life: 13 days.
Time to peak: About 3 days after a single 100 mg dose.
Excretion: In urine, mainly as unchanged drug.

Special Considerations

In Women: Not for use in women.
In Elderly Subjects: No substantial differences in safety relative to younger adults.

Abatacept**Brand Names****Europe**

Austria: Orenzia; **Belgium:** Orenzia; **Czech Republic:** Orenzia; **Denmark:** Orenzia; **Estonia:** Orenzia; **Finland:** Orenzia; **France:** Orenzia; **Germany:** Orenzia; **Greece:** Orenzia; **Hungary:** Orenzia; **Ireland:** Orenzia; **Italy:** Orenzia; **Latvia:** Orenzia; **Luxembourg:** Orenzia; **Netherlands:** Orenzia; **Poland:** Orenzia; **Portugal:** Orenzia; **Romania:** Orenzia; **Slovakia:** Orenzia; **Spain:** Orenzia; **Sweden:** Orenzia; **UK:** Orenzia.

North America

Canada: Orenzia; **USA:** Orenzia.

Latin America

Argentina: Orenzia.

Chemistry

Abatacept: (C₁₉₆₅H₃₀₈₀N₄₇₉O₆₉₅S₁₆). Mw: approx. 92000.01 (46000.01 per chain). 1-25-Oncostatin M (human precursor) fusion protein with CTLA-4 (antigen)(human) fusion protein with immunoglobulin G₁ (human heavy chain fragment). CAS-332348-12-6 (2004).

MGVLLTQRTL	LSLVLLALLFP	SMASAMMHVA	QPAVLASSR	GIASFVCEYA
SPGKATEVRV	TVLRQADSQV	TEVCAATYMM	GNELTFLDSD	ICTGTSSGNQ
VNLTIQGLRA	MDTGLYICKV	ELMYPPPYL	GIGNGIQIYV	IDPEPCPDS
QEPKSSDKTH	TSPSPAPPEL	LGSSSVFLFP	PKPKDTLMIS	RTPEVTCVVV
DVSHEDPEVK	FNWYVDGVEV	HNAKTKPREE	OYNSTYRVS	VLTVLHQDWL
NGKEYKCKVS	NKALPAIEK	TISKAKGQPR	EPQVYTLPPS	RDELTKNQVS
LTCLVKGFPY	SDIAVEWESN	GQFENNYKTT	PPVLDSGGSF	FLYSKLTVDK
SRWQQGNVFS	CSVMHEALHN	HYTKQSLSL	PG**K*	

** -C-terminus (predominant species)

* -C-terminus (cDNA)

Underlined: -N-glycosylation

Pharmacologic Category

Antirheumatic Drugs. (ATC-Code: L04AA24).

Mechanism of Action

Blocks CD28 interaction between antigen-presenting cells and T cells. Decreases T-cell proliferation and inhibits production of TNF, interferon-gamma, and interleukin-2. Suppresses inflammation, decreases anti-collagen antibody production, and reduces antigen-specific production of interferon gamma.

Therapeutic Use

Treatment of adult rheumatoid arthritis (as monotherapy or in combination with other disease-modifying anti-rheumatic drugs) or juvenile idiopathic arthritis (as monotherapy or in combination with methotrexate).

Pregnancy and Lactation Implications

Should be used during pregnancy only if clearly needed (risk of autoimmune disease in fetus). Not recommended during lactation.

Contraindications

Hypersensitivity to abatacept or any component of the formulation.

Warnings and Precautions

Hypersensitivity, anaphylaxis or anaphylactoid reactions might occur. May affect defenses against malignancies. Risk of lymphoma. Caution with chronic obstructive pulmonary disease (COPD). Screening for viral hepatitis prior to use. Live vaccines should not be given concurrently or within 3 months of discontinuation of therapy.

Adverse Reactions

Cardiovascular: Hypertension, hypotension.

Central Nervous System: Dizziness, headache.

Dermatologic: Cellulitis, pruritus, rash, urticaria.

Gastrointestinal: Abdominal pain, diarrhea, diverticulitis, dyspepsia, nausea.

Genitourinary: Pyelonephritis, urinary tract infection.

Hematologic: Acute lymphocytic leukemia.

Neuromuscular and Skeletal: Back pain, limb pain.

Respiratory: Bronchitis, COPD exacerbation, cough, dyspnea, lung cancer, nasopharyngitis, pneumonia, rhinitis, rhonchi, sinusitis, upper respiratory tract infection, wheezing.

Miscellaneous: Anaphylaxis, anaphylactoid reactions, antibody formation, disease flare, fever, flushing, herpes simplex and influenza infections, hypersensitivity, infusion-related reactions, joint wear, lymphoma, malignancies (including bile duct, bladder, breast, cervical, melanoma, myelodysplastic syndrome, prostate, renal, skin, thyroid and uterine), ovarian cyst, varicella infection.

Pharmacogenetics

Genes that may be involved:

CD28; CD80; CD86; CTLA4; IFNG; IL2; TNF

Drug Interactions

Should not be used in combination with anakinra or TNF-blocking agents (risk of infections). Live vaccines should not be administered during treatment or for 3 months following completion of the same. Concomitant use with SSRIs, lithium, valproic acid and carbamazepine may produce additive effects.

Nutrition/Nutraceutical Interactions

Herbs/Nutraceuticals: Avoid echinacea (immunostimulant properties).

Dosage

Dosage Forms: Injection, powder for reconstitution: 250 mg (contains maltose). [I.V. administration].

Pharmacokinetics and Pharmacodynamics

Absorption: Following multiple I.V. infusions, steady-state serum concentrations reached at 60 days. Systemic accumulation does not appear to occur.

Distribution: V_d: 0.02-0.13 L/kg.

Half-life: 8-25 days.

Special Considerations

In Elderly Subjects: Use with caution (higher incidence of infections and malignancies).

Monitoring Parameters: Signs and symptoms of infection and infusion reaction, hepatitis and tuberculosis screening prior to therapy initiation.

Abciximab**Brand Names****Europe**

Austria: Reopro; **Belgium:** Reopro; **Bulgaria:** Reopro; **Czech Republic:** Reopro; **Denmark:** Reopro; **Finland:** Reopro; **France:** Reopro; **Germany:** Reopro; **Greece:** Reopro; **Hungary:** Reopro; **Ireland:** Reopro; **Italy:** Reopro; **Luxembourg:** Reopro; **Netherlands:** ReoPro; **Poland:** ReoPro; **Portugal:** ReoPro; **Romania:** ReoPro; **Slovenia:** Reopro; **Spain:** Reopro; **Sweden:** Reopro; **UK:** Reopro.

North America

Canada: Reopro; **USA:** Reopro.

Latin America

Brazil: Reopro; **Mexico:** Reopro.

Chemistry

Abciximab: Mw: approx. 47600.01. (1) Immunoglobulin G₁, (human-mouse monoclonal c7E3 clone p7E3VhCg1 Fab fragment anti-human glycoprotein IIb/IIIa receptor), disulfide with human-mouse monoclonal c7E3 clone p7E3VhCk light chain; (2) Immunoglobulin G₁ (human-mouse monoclonal c7E3 clone p7E3VhCg1 Fab fragment anti-human glycoprotein IIb/IIIa receptor), disulfide with human-mouse monoclonal c7E3 clone p7E3VhCk light chain. CAS-143653-53-6 (1994).

Pharmacologic Category

Antithrombotic Agents; Platelet-aggregation Inhibitors; Glycoprotein IIb/IIIa Inhibitor. (ATC-Code: B01AC13).

Mechanism of Action

Fab antibody fragment of the chimeric human-murine monoclonal antibody 7E3. Inhibits platelet aggregation by binding to IIb/IIIa receptors.

Therapeutic Use

Prevention of acute cardiac ischemic complications in patients at high risk for abrupt closure of the treated coronary vessel and patients at risk of restenosis. Adjunct with heparin to prevent cardiac ischemic complications in patients with unstable angina not responding to conventional therapy when a percutaneous coronary intervention is scheduled within 24 hours.

Unlabeled Use

Myocardial infarction. Acute ischemic stroke.

Pregnancy and Lactation Implications

Not known whether fetal harm can be caused or if reproduction capacity can be affected. Caution during lactation.

Contraindications

Hypersensitivity. Active internal hemorrhage or recent GI or GU bleeding. History of cerebrovascular accident. Clotting abnormalities or recent administration of oral anticoagulants (within 7 days). Thrombocytopenia. Recent major surgery or trauma. Intracranial tumor. Arteriovenous malformation. Aneurysm. Severe uncontrolled hypertension. Vasculitis. Use of dextran before percutaneous transluminal coronary angioplasty. Concomitant use of another parenteral GP IIb/IIIa inhibitor.

Warnings and Precautions

Anaphylaxis/hypersensitivity reactions, thrombocytopenia as well as diminished efficacy might occur due to human antichimeric antibody formation. Bleeding complications (use with extreme caution in platelet counts <150,000/mm³, hemorrhagic retinopathy, previous history of gastrointestinal disease, recent thrombolytic therapy and in chronic dialysis. Use caution with administration of other drugs affecting hemostasis. Diminished drug efficacy (human antichimeric antibody formation). Use with caution in patients weighing <75 kg or >65 years of age (increased risk of bleeding).

Adverse Reactions

Cardiovascular: Arteriovenous fistula, bradycardia, chest pain, complete AV block, embolism, hypotension, incomplete AV block, nodal arrhythmia, palpitation, peripheral edema, pseudoaneurysm, stroke, thrombophlebitis, ventricular tachycardia.

Central Nervous System: Abnormal thinking, abnormal vision, agitation, anxiety, coma, confusion, diplopia, dizziness, headache, intracranial hemorrhage.

Dermatologic: Bullous eruption, cellulites, increased diaphoresis, petechiae, pruritus.

Endocrine and Metabolic: Diabetes mellitus, hyperkalemia.

Gastrointestinal: Abdominal pain, diarrhea, dyspepsia, gastroesophageal reflux, ileus, nausea, vomiting, xerostomia.

Genitourinary: Dysuria, prostatitis.

Hematologic: Anemia, leukocytosis, minor and major bleeding, thrombocytopenia.

Local: Injection site pain.

Neuromuscular and Skeletal: Back pain, hypertonia, muscle contractions, myalgia, weakness.

Respiratory: Bronchitis, bronchospasm, pleural effusion, pneumonia, pulmonary embolism.

Renal: Urinary incontinence, urinary retention.

Miscellaneous: Allergic reactions/anaphylaxis, cystalgia, inflammation, pain, peripheral coldness.

Pharmacogenetics

Caution and personalized dose adjustment in patients with the following genotypes:

ITGB3: Ex.2/Lau33Pro

Other genes that may be involved:

CYP1A2; CYP2C19; ITGA2; P2RY1

Drug Interactions

Antiplatelet agents may enhance the anticoagulant effect of anticoagulants. Antiplatelet agents and dasatinib may enhance the anticoagulant effect of other antiplatelet agents. Dextran may enhance the anticoagulant effect of abciximab. Antiplatelet agents may enhance the adverse/toxic effect of drotrecogin alpha (risk of bleeding), and of ibritumomab (impaired platelet function and risk of bleeding). Abciximab may enhance the potential for allergic or hypersensitivity reactions to monoclonal antibodies (thrombocytopenia or diminished therapeutic effects. Exceptions: alefacept). Nonsteroidal anti-inflammatory agents may enhance the adverse/toxic effect of antiplatelet agents (risk of bleeding). ω-3-Acid ethyl esters, and prostacyclin analogs may enhance the antiplatelet effect of antiplatelet agents. Pentosan polysulfate sodium may enhance the adverse/toxic effect of antiplatelet agents (risk of bleeding). Antiplatelet agents may enhance the adverse/toxic effect of salicylates (risk of bleeding). Antiplatelet agents may enhance the adverse/toxic effect of tositumomab and iodine ¹³¹I tositumomab (bleeding-related adverse events).

Nutrition/Nutraceutical Interactions

Herbs/Nutraceuticals: Herbs with anticoagulant/antiplatelet properties (e.g. alfalfa, anise, bilberry) may enhance the adverse/toxic effect of antiplatelet agents (risk of bleeding).

Dosage

Dosage Forms: Injection, solution: 2 mg/mL (5 mL). [I.V. administration].

World Guide for Drug Use and Pharmacogenomics

Genes

A

A2M (alpha-2-macroglobulin)

Alternative Names

Alpha-2M; C3 and PZP-like alpha-2-macroglobulin domain-containing protein 5.

Alternative Symbols

A2MG, CPAMD5, DKFp779B086, FWP007, S863-7.

Codes

OMIM: 103950. PharmGKB: PA24357.

Gene

Locus: 12p13.31.

Size: 48.44 kb, 36 Exons.

RNA

Transcripts/Variants: A2M-001: 4844 bp. A2M-002: 623 bp. A2M-003: 533 bp. A2M-004: 533 bp. A2M-005: 692 bp. A2M-006: 546 bp. A2M-007: 729 bp. A2M-008: 509 bp.

Expression: **Organs:** Liver. **Tissues:** Hematopoietic tissue. **Fluids:** Blood, Plasma. **Subcellular localization:** Extracellular.

Protein

Size: A2M-001: 163.3 kDa, 1474 aa. A2M-002: 18.72 kDa, 168 aa.

Family: Protease inhibitor I39 (α_2 -macroglobulin) family.

Category: Transport carrier.

Motifs/domains: Peptide stretch, called "bait region" with specific cleavage sites for different proteinases.

Function

A protease inhibitor and cytokine transporter. Inhibits many proteases, including trypsin, thrombin and collagenase. A2M is implicated in Alzheimer's disease (AD) due to its ability to mediate clearance degradation of A β , the major component of β -amyloid deposits. It is able to inhibit all four classes of proteinases by a unique "trapping" mechanism. This protein has a peptide stretch, called "bait region", which contains specific cleavage sites for different proteinases. When a proteinase cleaves bait region, a conformational change is induced in protein which traps proteinase. The entrapped enzyme remains active against low molecular weight substrates (activity against high molecular weight substrates greatly reduced). Following cleavage in bait region, a thioester bond is hydrolyzed and mediates covalent binding of protein to proteinase. A2M is a serum protease inhibitor mediating clearance and degradation of A β , the major component of β -amyloid deposits; a protease inhibitor and cytokine transporter. Associates with TGF β s and forms latent complexes; associates with PDGFV β ; has endopeptidase inhibitor activity; serine-type endopeptidase inhibitor activity; protein binding; wide-spectrum protease inhibitor activity; enzyme binding.

Related Diseases

Acne vulgaris; Acute kidney failure; Acute liver failure; Acute-phase reaction; Adenocarcinoma; Adrenal gland neoplasms; Adrenocortical adenoma; **Alzheimer's disease**; Amyloidosis; Arteriosclerosis; Autistic disorder; Autoimmune diseases; **Blood protein disorders**; Bone marrow neoplasms; Brain injuries; Breast neoplasms; Cardiovascular disease; Cataract; Cholangiocarcinoma; Cholangitis; Cholestasis; Colonic neoplasms; Colorectal neoplasms; Dementia; Diabetic neuropathy; Dyslipidemias; Edema; Endometriosis; **Emphysema due to A2M deficiency**; Esophageal neoplasms; Essential tremor; Ewing's sarcoma; Experimental arthritis; Experimental autoimmune encephalomyelitis; Experimental liver cirrhosis; Experimental liver neoplasms; Experimental mammary neoplasms; Fat necrosis; Fatty liver; Fever; Female genital neoplasms; Female infertility; Head and neck neoplasms; Hemolytic-uremic syndrome; Hepatic encephalopathy; Hepatitis; Hepatocellular carcinoma; Hodgkin's disease; Hyperalgesia; Hypospadias; **Inflammation**; Intrahepatic cholestasis; Iron metabolism disorders; Leishmaniasis; Liver diseases; Liver neoplasms; Lung neoplasms; Macular degeneration; Male infertility; Melanoma; Amelanotic melanoma; Metabolic syndrome; Multiple myeloma; Multiple sclerosis; *Mycobacterium* infections; Myocardial infarction; Neoplasms; **Nephrotic syndrome**; Neural tube defects; Niemann-Pick disease, type C; Non-small cell lung carcinoma; Obesity; **Obstructive lung diseases**; Osteoporosis; Ovarian neoplasms; Pain; Pancreatic neoplasms; Panic disorder; Parkinson's disease; Pneumonia; Polycystic ovary syndrome; Prostatic neoplasms; Pruritus; Adult respiratory distress syndrome; Respiratory hypersensitivity; **Rheumatoid arthritis**; Salivary gland neoplasms; Seizures; Skin neoplasms; Spermatocoele; Spinal cord injuries; Spinal dysraphism; Squamous cell carcinoma; Stomach diseases; Stomach neoplasms; Stomach ulcer; Testicular neoplasms; Thrombophilia; Thrombosis; Toxic hepatitis; Traumatic brain hemorrhage; Trypanosomiasis; Tuberculosis; Urinary bladder neoplasms; Uterine cervical neoplasms; Uterine neoplasms; Vaginal neoplasms; Vascular diseases; Venous thrombosis; Vomiting.

Related Drugs

1-Naphthylisothiocyanate; 3,4-Dichloroaniline; Acetaminophen; Antimony; Antimony potassium tartrate; Bacitracin; Becaplermin; beta-Naphthoflavone; Cisplatin; Clofibrate; Clofibrate; Clofibrate; Clofibrate; Corticosterone; Dexamethasone; Dimethylnitrosamine; Dimethyl sulfoxide; Ethinylestradiol; Ethinyl estradiol-desogestrel combination; Fluticasone propionate; Furan; Glucocorticoids; Imidazole; Imidazole-bis(imidazole)dimethylsulfoxideimidazotetrachlororuthenate(III); Isoniazid; Lipopolysaccharides; Mercury; Methylphenylamine; Naphthalene; Nitrosobenzylmethylamine; Pirinixic acid; Progesterone; Sodium arsenite; Thioacetamide; Turpentine.

Animal Models

Chimeric mouse. Herring and coworkers identified the up-regulation of *a2m* in TgCRND8 mice (carrying human APP (Swedish/Indiana)) by affecting A β receptor/transporter molecules, facilitating A β clearance by brain vascularization. The up-regulation of *a2m* was also investigated by Petrusz and colleagues in transgenic mice that overexpressed rat androgen-binding protein

(ABP) in their testes, resulting in a spermatogenesis impairment. The up-regulated *a2m* in these transgenic mice seemed to mediate a testicular response to the reduced levels of androgens and therefore, to modulate the spermatogenesis deficit.

Transgenic mice. *A2m* knockout mice are not only viable, but more resistant to endotoxin. They produce normal-sized litters and show no obvious phenotypic abnormalities. Murine *a2m* binds TGF- β and inhibits TGF- β -receptor interactions, which might explain the endotoxin-insensitive phenotype of knockout mice.

A2M-deficient (*MAM*^{-/-}) mice were reported by Hochejied and colleagues to be significantly protected against lethal systemic inflammation induced by the tumor necrosis factor (TNF). As a result, the MAM deficiency shows a high metabolic tolerance to TNF, being essential for studying the pathogenesis of Gram-negative septic shock. Also using this type of transgenic mice (*MAM*^{-/-}), Umans et al defined *in vivo* the role of macroglobulins in pancreatitis by analyzing the clinical symptoms and the expression of different cytokines and polypeptide factors. Once acute pancreatitis was induced in A2M-deficient mice, severe clinical symptoms and high mortality were observed, together with the significant induction of interleukin-1-receptor antagonist mRNA and most polypeptide factors (tumor necrosis factor- α , tumor necrosis factor- β , beta-lymphotoxin, and interferon- γ mRNA).

Allelic Variants

Total variants: 378.

Selected SNPs

rs669. HGVS: NM_000014.4: c.2998A>G; NP_000005.2: p.I1000V; NT_009714.16: g.1991241T>C. Susceptibility to Alzheimer's disease. A sequence polymorphism near the thioester site of the gene, changing Val1000 (GTC) to Ile (ATC); the 2 alleles had frequencies of 0.30 and 0.70, respectively. No difference of A2M serum levels observed for these 2 alleles. Increased frequency of G/G genotype from 0.07 in controls to 0.12 in AD. Synergistic effect with a myeloperoxidase promoter polymorphism genotype (-463G/G). Alleles: African: A: 0.658, G: 0.342; Asian: A: 0.875-0.967, G: 0.033-0.125; European: A: 0.658-0.750, G: 0.250-0.342; Multi-national: A: 0.778, G: 0.222. Genotypes: African: A/A: 0.467, A/G: 0.383, G/G: 0.150; Asian: A/A: 0.795-0.933, A/G: 0.067-0.159, G/G: 0.000-0.045; European: A/A: 0.417-0.625, A/G: 0.250-0.483, G/G: 0.100-0.125; Multi-national: A/A: 0.741, A/G: 0.074, G/G: 0.185.

rs226379. HGVS: NT_009714.16: g.2027559T>C. Associated with Alzheimer's disease. Alleles: African: A: 0.593, G: 0.407; Asian: A: 0.867-0.935, G: 0.065-0.133; European: A: 0.630-0.750, G: 0.250-0.370; North American: A: 0.543-0.625, G: 0.375-0.457. Genotypes: African: A/A: 0.356, A/G: 0.475, G/G: 0.169; Asian: A/A: 0.778-0.913, A/G: 0.043-0.178, G/G: 0.043-0.044; European: A/A: 0.370-0.625, A/G: 0.250-0.522, G/G: 0.109-0.125; North American: A/A: 0.348-0.375, A/G: 0.391-0.500, G/G: 0.125-0.261.

rs226380. HGVS: NM_000014.4: c.-28T>G; NT_009714.16: g.2027447A>C. Alleles: African: G: 0.850, T: 0.150; Asian: G: 0.133-0.167, T: 0.833-0.867; European: G: 0.438-0.458, T: 0.542-0.562; Multi-national: G: 0.130, T: 0.870; North American: G: 0.848, T: 0.152. Genotypes: African: G/G: 0.700, G/T: 0.300, T/T: 0.000; Asian: G/G: 0.022-0.067, G/T: 0.200-0.222, T/T: 0.733-0.756; European: G/G: 0.217-0.250, G/T: 0.375-0.483, T/T: 0.300-0.375; North American: G/G: 0.696, G/T: 0.304, T/T: 0.000.

rs226405. HGVS: NM_000014.4: c.1915G>A; NP_000005.2: p.Asp639Asn; NT_009714.16: g.2007207T>C. Alleles: African: A: 0.017, G: 0.983; Asian: A: 0.000, G: 1.000; European: A: 0.000, G: 1.000; Multi-national: A: 0.000, G: 1.000. Genotypes: African: A/A: 0.033, G/G: 0.967; Asian: A/G: 0.000, G/G: 1.000; European: A/G: 0.000, G/G: 1.000; Multi-national: A/G: 0.000, G/G: 1.000.

rs3026229. HGVS: NT_009714.16: g.1978874G>T. Alleles: African: A: 0.195, C: 0.805; Asian: A: 0.022-0.065, C: 0.935-0.978; European: A: 0.089-0.188, C: 0.812-0.911; North American: A: 0.205, C: 0.795. Genotypes: African: A/A: 0.051, A/C: 0.288, C/C: 0.661; Asian: A/A: 0.000-0.043, A/C: 0.043-0.044, C/C: 0.913-0.956; European: A/A: 0.000, A/C: 0.179-0.375, C/C: 0.625-0.821; North American: A/A: 0.045, A/C: 0.318, C/C: 0.636.

rs10771309. HGVS: NT_009714.16: g.1977580C>A. Alleles: Asian: A: 0.033-0.111, C: 0.889-0.967; European: A: 0.342, C: 0.658. Genotypes: African: A/A: 0.000-0.044, A/C: 0.067-0.133; European: A/A: 0.100, A/C: 0.483.

rs12309184. HGVS: NT_009714.16: g.2027808G>A. Alleles: African: A: 0.025, G: 0.975; Asian: A: 0.000, G: 1.000; European: A: 0.000, G: 1.000. Genotypes: African: A/G: 0.050, G/G: 0.950; Asian: A/G: 0.000, G/G: 1.000; European: A/G: 0.000, G/G: 1.000.

rs1800433. HGVS: NM_000014.4: c.2915G>A; NP_000005.2: p.C972Y; NT_009714.16: g.1991324C>T. A mutation within the thioester site, changing Cys972 (TGT) to Tyr (TAT). Since activation of the internal thioester formed between Cys972 and Gln975 in each of the subunits of the tetrameric A2M molecule is involved in the covalent crosslinking of the activating proteinase, this mutation was predicted to interfere with A2M function; however, A2M serum levels are within normal range in mutants.

rs1805667. HGVS: NT_009714.16: g.1978940A>C. Alleles: African: G: 0.583, T: 0.417; Asian: G: 0.062-0.111, T: 0.889-0.938; European: G: 0.250-0.342, T: 0.658-0.750; North American: G: 0.591, T: 0.409. Genotypes: African: G/G: 0.350, G/T: 0.467, T/T: 0.183; Asian: G/G: 0.044-0.042, G/T: 0.042-0.133, T/T: 0.822-0.917; European: G/G: 0.100-0.125, G/T: 0.250-0.483, T/T: 0.417-0.625; North American: G/G: 0.318, G/T: 0.545, T/T: 0.136.

rs1805688. HGVS: NT_009714.16: g.1979074A>G. Alleles: African: C: 0.092, T: 1.000; Asian: C: 0.000, T: 1.000; European: C: 0.000, T: 1.000. Genotypes: African: C/T: 0.183, T/T: 0.817; Asian: C/C: 0.000, T/T: 1.000. European: C/C: 0.000, T/T: 1.000.

rs3759277. HGVS: NG_011717.1: g.52360C>T; NM_000014.4: c.4366+137C>T; NT_009714.17: g.1981323G>A. Alleles: African: C: 0.183, T: 0.817; Asian: C: 0.875, T: 0.125; European: C: 0.542, T: 0.458; North American: C: 0.159, T: 0.841. Genotypes: African: C/C: 0.017, C/T: 0.333, T/T: 0.650; Asian: C/C: 0.833, C/T: 0.083, T/T: 0.083; European: C/C: 0.300, C/T: 0.483, T/T: 0.217; North American: C/C: 0.000, C/T: 0.318, T/T: 0.682.

lvs1Del: A deletion of the intron that separates exons 1 and 2, resulting in fusion of the two exons that code the bait domain of the *A2M* gene.

Arg681His: A G-to-A transition in exon 17, predicting an Arg-to-His substitution at position 681. In the mutant allele a MaeII restriction site was lost and a new NspHI site was created.

Ex18Del: Susceptibility to Alzheimer's disease. A 5'-splice site deletion in exon 18 (*A2M-2*). This exon encodes 'exon II' of the bait domain of *A2M*, which functions to attract and trap proteases. The *A2M-2* deletion confers increased risk for Alzheimer's disease.

5-bp Del: A deletion of 5 bases from -7 to -3 from the 5' splice site of exon 2.

Evolution

Human α_2 -macroglobulin, complement component C3 and complement component C4 contain a unique activatable β -cysteinyl- γ -glutamyl thiol ester and have a common evolutionary origin. C5 also shows sequence homology to *A2M*.

Genomics and Pharmacogenomics

Alzheimer's disease: *A2M* has been recognized as a candidate gene for late-onset Alzheimer's disease (AD), but the association between several polymorphisms in the *A2M* gene and risk for AD remained controversial. Moreover, little is known regarding the effects of polymorphisms in the *A2M* promoter region on AD susceptibility. Song et al set out to detect polymorphisms in the *A2M* promoter region, and then evaluate their relationship to sporadic AD (SAD). One SNP (-88A/G) in the proximal promoter region was found by sequencing, and further analyzed with an established 25T/G polymorphism in 179 SAD patients and 179 age-gender-matched controls. Allele A in the -88A/G polymorphism was more prevalent in cases, with a 1.7-folded risk for SAD, while the G allele in 25T/G was less prevalent in cases, with a 43% reduced risk for SAD. After adjusting the effects of age, gender and APOE- ϵ 4 allele status in a logistic regression model, the protective effects of -88G and 25G on SAD still remained. Individuals who carried the haplotype -88G/25G had a significant 44% reduced risk for SAD compared to those who did not carry it, while haplotype -88A/25T carriers had an increased risk for SAD compared to those who did not carry it. This study supports the hypothesis that haplotype -88G/25G might play a protective role in the development of SAD, and the protective effects of -88G and 25G were independent of APOE- ϵ 4 allele.

A2M is a highly plausible candidate gene for Alzheimer's disease (AD) in a region of chromosome 12 which has numerous independent reports of genetic linkage. A 5 bp deletion in *A2M* associated with AD in a subset of the National Institute of Health (NIMH) Genetics Initiative AD family sample. Efforts to replicate this association in case-control samples were largely negative, while those in family samples were more positive. Variable findings regarding this deletion, along with variable reports of association with V1000I, another polymorphism in the gene, result from linkage disequilibrium in the area as well as ascertainment differences between family-based and case-control studies. Seven novel polymorphisms were identified in the full NIMH sample of 1439 individuals in 437 families. There is genetic association of the 5 bp deletion and two novel polymorphisms with AD. Substantial linkage disequilibrium was detected across the gene as a whole, and haplotype analysis also showed significant association between AD and groups of *A2M* polymorphisms. Several of these polymorphisms and haplotypes remain significantly associated with AD even after correction for multiple testing.

A2M, the *A2M* receptor (LRP1), two low density lipoprotein-related protein (LRP) ligands, APOE, APP, BACE, BLMH, PS1, and PS2 are probably genetically linked in regulation of brain amyloidogenesis. *A2M* is a carrier protein for β -amyloid decreasing fibril formation and influencing amyloid neurotoxicity. *A2M* enhances clearance of β -amyloid via low-density lipoprotein receptor-related protein in primary Tg2576 transgenic mouse cortical neurons. In recent years, there have been over 100 genetic studies in different countries debating the controversial role of a five-base-pair intronic deletion in the *A2M* gene in LOAD. The sibship disequilibrium test revealed a significant association between *A2M* and AD; and the inheritance of a deletion in the *A2M* gene at the 5'-splice site of exon II of the bait region (exon 18)(*A2M-2*) confers increased risk for AD, but these findings could not be replicated by others in different populations and races, though in Koreans the *A2M-D* allele might represent a modest risk factor for LOAD. The *A2M-V1000I* polymorphism (GTC/ATC) was also found to be associated with AD, but no association was found between the *A2M* polymorphisms (intronic 5-bp deletion, Ile1000Val) and AD. *A2M* G2998A mutation is not associated with AD in the Chinese population. In addition, *A2M* is not associated with age at onset, duration of illness, senile plaques, dystrophic neurites or NFT in AD. No change in *A2M* mRNA, protein, or protein expression could be found in AD and it appears that *A2M* is not genetically associated with LOAD. In Spanish patients, the *A2M-G/G* genotype is more frequent in AD (8.1%) than in controls (3.79%), and the homozygous *A2M* deletion (*A2M-D/D*) is absent in controls and appears in 5.21% of AD patients. The integration of *A2M*-related polymorphic variants in single clusters revealed clear differences in the distribution of the *A2M* genotypes between AD and controls. Data indicate that the *A2M* gene polymorphism can confer an increased risk for AD with an estimated Mantel-Haenszel ratio of 1.5 and no age- or gender-dependent increase in *A2M* gene allele frequencies. Plasma from patients homozygous for the *A2M* deletion show normal *A2M* subunit size, conformation, and proteinase inhibitory activity. In some cases, *A2M* exhibits a marked increase in TGF β 1 binding. Methylamine-treated *A2M* mutant samples show modest elevations in β -amyloid binding to *A2M* compared with samples from patients lacking the deletion, suggesting a possible functional basis by which *A2M* deletion may influence multifactorial AD pathogenesis.

Genetic association with LOAD susceptibility studied in 738 Caucasian families (4704 individuals) and an independent case-control dataset with 296 cases and 566 controls explored 11 candidate genes (47 SNPs common to both samples). In addition to tests for main effects and haplotypes, MDR-PDT was used to search for gene-gene interactions in the family data. Significant haplotype effects in *ACE* in family and case-control samples using standard and cladiatic haplotype models were observed. *ACE* was also part of significant 2 and 3-locus MDR-PDT joint effects models with *A2M*, which mediates the clearance of β -amyloid, and lucine-rich repeat transmembrane-3 (*LRRTM3*), a nested gene in α -3catenin (*CTNNA3*) which binds presenilin-1. These genes are related to β -amyloid clearance; thus this constellation of effects might constitute an axis of susceptibility for LOAD.

A panel of 15 proteins, including complement factor-H (CFH) and *A2M*, validated as AD-specific plasma biomarkers. Both CFH and *A2M* show positive correlations with hippocampal metabolite ratio N-acetylaspartate/myo-inositol (NAA/mI) in AD. Association of plasma CFH and *A2M* with hippocampal NAA/mI in AD subjects suggests that these proteins may reflect disease progression in early AD.

Fluticasone propionate: In patients with asthma, fluticasone induces reduction in *A2M* plasma levels. No pharmacogenetic data are available.

Glucocorticoids: A detailed time course of the assembly and disassembly of a STAT3-dependent, glucocorticoid-supplemented enhanceosome for the *A2M* gene has been described and compared with a detailed time course of transcription of the gene by run-on analysis. The glucocorticoid receptor (GR) can associate with the enhanceosome without STAT3. The enhanceosome contains c-Jun/c-Fos and OCT-1 constitutively. All these factors (GR, c-Jun, OCT-1) have transcription activation domains, but STAT3 is required for the observed transcriptional increase. The time course of enhanceosome occupation by GR and tyrosine-phosphorylated STAT3 shows that these transcription factors precede the arrival of RNA polymerase II (Pol II) by approximately 5-10 min. The enhanceosome remains assembled for approximately 90 min in the continued presence of both inducers. When IL6 and dexamethasone are removed (after 30 min of treatment), the disappearance within an additional 30 min of the established enhanceosome indicates that renewal of STAT3 and GR binding must occur in the continued presence of IL6+Dex. Compared with the total nuclear tyrosine-phosphorylated STAT3 capable of binding DNA, the chromatin-associated STAT3 resists dephosphorylation and appears to recycle to maintain the enhanceosome. Run-on transcription shows a lag after full enhanceosome occupation that can be largely but not completely explained by the approximately 30 min transit time of Pol II across the *A2M* locus.

Inflammation: Acute-phase protein synthesis in liver during inflammation is regulated via cytokines and glucocorticoids. Using quantitative reverse transcription (RT)-PCR analysis and immunoassay, it has been shown that the acute-phase protein, *A2M*, is activated after systemic inflammation induced by lipopolysaccharide (LPS) or localized inflammation induced by turpentine oil (TO). Addition of heme during LPS-induced inflammation can boost the expression of *A2M*, whereas blocking heme synthesis (by succinyl acetone) or enhancing its consumption in parallel biosynthetic pathways (cytochrome P450 induction by phenobarbital) decreases *A2M* expression. This decrease is abolished by exogenous heme supplementation. Heme supplementation is also able to increase *A2M* response in female rats to a level similar to that in male rats providing a new insight into the puzzling sexual dimorphism observed previously during localized inflammation. Heme might be considered a regulatory element in controlling liver *A2M* expression during inflammation.

Nephrotic syndrome: In both nephrotic syndrome (NS) and hereditary analbuminemia in the Nagase analbuminemic rat (NAR), the plasma protein concentration is nearly normal since albumin is replaced by several high molecular weight proteins. In rats these include protease inhibitors *A2M*, a 720 kDa positive acute phase protein (APP) and α_2 -inhibitor 3 (α_2 -I3), a 180 kDa negative APP. There is no known stimulus to increase α_2 -I3 synthesis, but like albumin and other negative APPs its synthesis decreases during inflammation by transcriptional down-regulation. In hypoalbuminemic states gene transcription of other positive and negative APPs is increased. *A2M* is increased significantly (12-fold) in NAR and by approximately 50-fold in rats with NS compared to control. The α_2 -I3 concentration is twice normal in NAR or NS compared to controls, providing approximately half of the total plasma protein. Hepatic *A2M* mRNA increases only 30% in NAR and twofold in NS, suggesting post-transcriptional regulation.

Rheumatoid arthritis: Genetic associations of *TNFR2*, *VDR* (*Bsm I* and *Fok I*), *A2M*, *GSTT1*, *GSTM1* and *ACE* in South Asian and Caucasian patients with rheumatoid arthritis (RA) were assessed in a study by Ghelani et al. DNA samples from South Asians (134 cases, 149 controls) and Caucasians (137 cases, 150 controls) from the East Midlands of the United Kingdom were genotyped for seven polymorphisms. All cases were rheumatoid-factor positive. Significant genetic associations were observed with *TNFR2* R>R, *A2M* 1-1 and *GSTT1*-null among Caucasian patients. In South Asians, *VDR Bsm I* B-B genotype, *A2M* 2-2 genotype, and *GSTT1*-null genotype were associated with RA. In the majority of cases, recessive and multiplicative modes of inheritance explained the observed associations. The study demonstrated that ethnicity affects the genetic associations in RA.

Drug-Gene Interactions

1-Naphthylisothiocyanate: Increases expression of *A2M* mRNA. **3,4-Dichloroaniline:** Decreases expression of *A2M* mRNA. **Acetaminophen:** Increases expression of *A2M* mRNA. **Antimony:** Decreases expression of *A2M* mRNA. **Antimony potassium tartrate:** Decreases expression of *A2M* mRNA. **beta-Naphthoflavone:** Decreases expression of *A2M* mRNA. **Cisplatin:** Binds to *A2M* protein. **Clofibrate:** Increases expression of *A2M* mRNA. **Clofibric acid:** Affects expression of *A2M* mRNA. **Corticosterone:** Increases expression of *A2M* mRNA. **Dexamethasone:** Decreases expression of *A2M* mRNA. **Dimethylnitrosamine:** Decreases expression of *A2M* mRNA. **Dimethyl sulfoxide:** Increases expression of *A2M* mRNA. **Ethinyl estradiol:** Decreases expression of *A2M* mRNA. **Ethinyl estradiol-desogestrel combination:** Increases expression of *A2M* protein. **Furan:** Decreases expression of *A2M* mRNA. **Imidazolium-bis(imidazole)di methylsulfoxideimidazotetetrachlororuthenate(III):** Binds to *A2M* protein. **Isoniazid:** Increases expression of *A2M* mRNA. **Lipopolysaccharides:** Increase expression of *A2M* mRNA. **Mercury:** Decreases expression of *A2M* mRNA. **Methapyriline:** Decreases expression of *A2M* mRNA. **Naphthalene:** Binds to *A2M* protein. **Nitrosobenzylmethylamine:** Increases expression of *A2M* mRNA. **Pinirixic acid:** Decreases expression of *A2M* mRNA. **Progesterone:** Affects expression of *A2M* mRNA. **Sodium arsenite:** Increases expression of *A2M* mRNA. **Thioacetamide:** Increases expression of *A2M* mRNA. **Turpentine:** Increases expression of *A2M* mRNA.

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ABCA1 (ATP-binding cassette, sub-family A (ABC1), member 1)

Alternative Names

ATP-binding cassette 1; ATP binding cassette transporter 1; ATP-binding cassette transporter A1; ATP-binding cassette, sub-family A member 1; ATP-binding cassette, sub-family A (ABC1), member 1; Cholesterol efflux regulatory protein; High density lipoprotein cholesterol level quantitative trait 13; High density lipoprotein deficiency, OTTHUMP00000021833; Tangier disease; Tangier type, 1.

Alternative Symbols

ABC-1, ABC1, CERP, FLJ14958, HDLCQ13, HDLDT1, MGC164864, MGC165011, TGD, Tangier.

Codes

OMIM: 600046. PharmGKB: PA24373.

Gene

Locus: 9q31.1.

Size: 147.15 kb, 50 Exons.

Regulatory sequence: Promoter (CAAT box), binding site. In the promoter region of *ABCA1* there are 7 putative SP1-binding sites, four sterol regulatory elements (SREs) similar to the SRE of the low density lipoprotein receptor (LDLR) promoter region, a CpG island, a possible weak TATA box, 2 distal CCAAT sequences, and binding sites for several other transcription factors.

Motifs/domains: Repetitive sequence ALU.

Structure: Numerous SP1 and putative sterol regulatory elements, binding sites for NFkB and activator proteins, conserved non coding sequences.

RNA

Transcripts/Variants: ABCA1-001: 1540 bp. ABCA1-002: 10494 bp. ABCA1-004: 923 bp. ABCA1-201: 1556 bp. ABCA1-003: 807 bp.

Expression: Organs: Brain, Liver, Mammary gland, Placenta, Prostate, Testis, Uterus (*Highly*); Adrenal gland, Kidney, Lung, Skin, Small intestine (*Moderately*); Bladder, Heart, Large intestine, Lymph node, Ovary, Pancreas, Spleen, Stomach, Thymus, Thyroid. **Tissues:** Bone marrow, Connective, Epithelial, Secretory, Striated/skeletal muscle (*Moderately*). **Cells:** Digestive, Leukocyte, Monocyte, Macrophage, Sertoli cell. **Subcellular localization:** Intracellular (cytoplasm, organelle, endosome); Plasma membrane.

Protein

Size: ABCA1-001: 41.12 kDa, 363 aa. ABCA1-002: 254.30 kDa, 2261 aa. ABCA1-004: 13.83 kDa, 123 aa. ABCA1-201: 13.64 kDa, 121 aa.

Family: ATP binding cassette superfamily; ABC transporter family; ABCA subfamily. Complete sequence of the yeast genome predicts the existence of 29 proteins belonging to the ubiquitous ATP-binding cassette (ABC) superfamily.

Using binary comparison, phylogenetic classification, and detection of conserved amino acid residues, the yeast ABC proteins have been classified in a total of 6 clusters, including 10 subclusters of distinct predicted topology and presumed distinct function. Study of the yeast ABC proteins provided insight into the physiologic function and biochemical mechanisms of their human homologs, such as those involved in cystic fibrosis, adrenoleukodystrophy, Zellweger syndrome, multidrug resistance, and the antiviral activity of interferons.

Category: Regulatory, transport carrier.

Motifs/domains: N terminal extracellular domain containing a segment homologous to the autoantigen SSN; two ATP binding (NBF) sites; two transmembrane (2TM) domains (2 x 6 segments) separated by a linker region; PEST sequence involved in internalization and cholesterol efflux from late endosomes; contains 2 highly conserved ATP-binding cassettes including Walker A and B motifs.

Post-translational modifications: Phosphorylation on Ser-2054 regulates phospholipid efflux.

Function

ABCA1 functions as a cholesterol efflux pump in the cellular lipid removal pathway. ABC1 is a key gatekeeper influencing intracellular cholesterol transport, named "cholesterol efflux regulatory protein" (CERP). ABCA1 binds ATP in the presence of Mg²⁺ and shows that ABCA1 expression supports apolipoprotein A-I (APOA1)-mediated release of cholesterol and choline-phospholipids. ABC transporter, traffic ATPase, facilitator of cellular efflux of cholesterol and phospholipid and key regulator of high density lipoprotein (HDL) metabolism; a leukocyte factor that controls the recruitment of inflammatory cells and protects against atherosclerosis; plays a key role in the first steps of the reverse cholesterol transport pathway by mediating lipid efflux from macrophages; plays an important role in lipid transport in Sertoli cells and influences male fertility; contributes to transport of maternal cholesterol to the developing fetus. Binding of APOA1 leads to the formation of phospholipid-APOA1 complexes which subsequently promote cholesterol efflux; binds to FADD, required for its function; carries cholesterol out of the cell for uptake into sterol dependent HDL particles regulated in monocytes and macrophages; inhibited by: SREBF2 in endothelial cells and PDGFA in vascular smooth muscle cells.

The membrane-associated protein encoded by this gene is a member of the superfamily of ATP-binding cassette (ABC) transporters. ABC proteins transport various molecules across extra- and intracellular membranes. ABC genes are divided into seven distinct subfamilies (ABCA1, MDR1/TAP, MRP, ALD, OABP, GGN20, White). This protein is a member of the ABC1 subfamily. Members of the ABC1 subfamily comprise the only major ABC subfamily found exclusively in multicellular eukaryotes. With cholesterol as its substrate, this protein functions as a cholesterol efflux pump in the cellular lipid removal pathway. Mutations in this gene have been associated with Tangier's disease and familial high-density lipoprotein deficiency. A cAMP-dependent and sulfonyleurea-sensitive anion transporter. A key gatekeeper influencing intracellular cholesterol transport. ABCA1 participates in the statin pathway (cholesterol and lipoprotein transport PD), PID pathways (RXR and RAR heterodimerization with other nuclear receptor), and reactome pathways (HDL-mediated lipid transport).

Related Diseases

Acute kidney failure; Acute myeloid leukemia; Acute promyelocytic leukemia; Adenoma; Alopecia; **Alzheimer's disease;** Analphalipoproteinemia; Animal mammary neoplasms; **Arteriosclerosis;** Asthma; **Atherosclerosis;** Autistic disorder; **Autosomal dominant hypercholesterolemia;** Barrett's esophagus; B-cell chronic lymphocytic leukemia; B-cell lymphoma; Blood coagulation disorders; Brain ischemia; Breast neoplasms; Bronchopulmonary dysplasia; Bullous pemphigoid; Cachexia; **Cardiovascular diseases;** Cataract; **Cerebral amyloid angiopathy;** Cervical intraepithelial neoplasia; Choriocarcinoma; **Chronic kidney disease;** **Chronic renal insufficiency;** Cleft lip; Cleft palate; Colitis; Colonic neoplasms; **Colorectal cancer;** **Colorectal neoplasms;** Congenital abnormalities; **Coronary heart disease;** **Coronary artery disease;** **Coronary disease;** Craniofacial abnormalities; Diabetes mellitus (type 2); Diabetic nephropathy; Diabetic neuropathy; Drug eruptions; Drug-induced abnormalities; **Dyslipidemias;** Edema; Embryonal carcinoma; Endometrial hyperplasia; Endometrial neoplasms; Epidermolysis bullosa; Epilepsy; Experimental arthritis; Experimental autoimmune encephalomyelitis; Experimental diabetes mellitus; Experimental liver cirrhosis; Experimental mammary neoplasms; Experimental neoplasms; Eye abnormalities; **Familial high density lipoprotein deficiency;** **Familial hypercholesterolemia;** Fatty liver; Fetal death; Fever; Fibrosis; Gingival hyperplasia; Gingival overgrowth; Glioblastoma; **HIV infection;** Head and neck neoplasms; **Heart diseases;** Hemolytic-uremic syndrome; Hepatocellular carcinoma; Hepatomegaly; Herpes simplex; **High density lipoprotein deficiency type 1 (HDL1); High density lipoprotein deficiency type 2 (HDL2);** Human influenza; Hyperalgesia; **Hyperalphalipoproteinemia;** **Hypercholesterolemia;** Hyper eosinophilic syndrome; Hyperhomocysteinemia; **Hyperlipidemia;** Hyperlipoproteinemia (type II); Hyperlipoproteinemia (type IIA); Hypertension; Hypoalphalipoproteinemia; Inflammation; Insulin resistance; Interstitial nephritis; Intestinal diseases; Iron metabolism disorders; Ischemic stroke; **Kidney diseases;** Learning disorders; Left ventricular dysfunction; Left ventricular hypertrophy; Leukemia; Lewis lung carcinoma; Leydig's cell tumor; Lipoprotein deficiencies; Lipoprotein disorder; Liver diseases; Liver neoplasms; Lung neoplasms; Medulloblastoma; Melanoma; Meningoencephalocele; Metabolic diseases; Middle cerebral artery infarction; Multiple myeloma; Muscular diseases; *Mycobacterium* infections; Myeloid leukemia; Myocardial infarction; Myocardial ischemia; Myocardial reperfusion injury; Myocarditis; Myositis; Myotonia congenita; Necrotizing enterocolitis; Neoplasms; Neurodegenerative diseases; Neurogenic inflammation; **Niemann-Pick type C disease;** Noise-induced hearing loss; Non-Hodgkin lymphoma; **Obesity;** Osteomalacia; Ovarian neoplasms; Pain; Pancreatic neoplasms; Partial epilepsies; Peritonitis; Pneumonia; Porokeratosis; Postmenopausal osteoporosis; Prenatal exposure delayed effects; Primary hypoalphalipoproteinemia; Prion diseases; Progressive myoclonic epilepsies; **Prostate cancer;** Prostatic hyperplasia; **Prostatic neoplasms;** Pseudolymphoma; Pterygium; Reperfusion injury; Respiratory hypersensitivity; Rhabdomyolysis; Rhabdomyosarcoma; Rheumatoid arthritis; **Scott syndrome;** Seborrheic keratosis; Skin neoplasms; **Smith-Lemli-Opitz syndrome;**

Smoking: Spinal muscular atrophy; Squamous cell carcinoma; Stomach neoplasms; **Stroke;** **Tangier disease;** **Thalidomide-related neuropathy;** Thyroid neoplasms; Tongue neoplasms; Toxic hepatitis; Tuberculosis; Uterine cervical neoplasms; Uterine neoplasms; Uveal neoplasms; Vitiligo; Wilms' tumor.

Related Drugs

1-Butanol; 1-Cleoyl-2-acetyl glycerol; 1-Palmitoyl-2-(5-oxovaleryl)-sn-glycero-3-phosphorylcholine; 4,4'-Diisothiocyanatostilbene-2,2'-disulfonic acid disodium salt; 9,11-Linoleic acid; 22-Hydroxycholesterol; 25-Hydroxycholesterol; AGN 193109; alpha-Lipoic acid; Alitretinoin; Arachidonic acid; Arsenic; Aspirin; Atorvastatin; ATI-111; Berberine; Calcitriol; Calmodulin; Charcoal-stripped fetal bovine serum; Cholesterol; Cholic acid; Chromium; Cilostazol; Daidzein; Dietary cholesterol; Dietary fats; Eicosapentaenoic acid; Ezetimibe; Fatty acids; Fenofibrate; Fluvastatin; Genistein; Ghrelin; *Ginkgo biloba* L.; Glyburide; Glycine; GSK 3987; GW 1929; GW 3965; HDL-cholesterol; Honokiol; HT31; Ibuprofen; Linoleic acid; Lipids; Lipopolysaccharides; Lithocholic acid; Liver X receptor agonists; Lovastatin; Low-density lipoproteins (LDL); Lycopene; Mastoparan; Methotrexate; Methyl-3β-hydroxy-5α,6α-epoxycholestanol (MHEC); miR-33; Mitoxantrone; Naringenin; N-(2,2,2-Trifluoro-ethyl)-N-[4-(2,2,2-trifluoro-1-hydroxy-1-trifluoromethyl-ethyl)-phenyl]-benzenesulfonamide (TO-901317); N-3-Oxo-dodecanoyl-Homoserine lactone (3OC12-HSL); Obestatin; Oleic acid; Ozone; Palmitoleic acid; *Panax notoginseng* saponins; Perfluorooctanoic acid; Phenylsulfone-substituted quinoxaline (WYE-672); Phenytoin; Phosphatidylcholine; Phosphodiesterase (PDE) type 4 inhibitors; Phospholipids; Pirinixic acid; Platelet Activating Factor; PPARG agonists; Pravastatin; Pregnenolone carbonitrile; Resveratrol; Retinoids; Reginoid LG268; Rifampin; Ritonavir; Rolipram; Rosiglitazone; Rosuvastatin; Simvastatin; Soy protein; Statins; Stigmasterol; Synthetic steroidal liver X receptor agonists; tert-Butylhydroperoxide; Tetracycline; Torcetrapib; trans-10,cis-12-Conjugated linoleic acid; Tretinoin; Triacsin C; Tributyltin chloride (TBTC); Vistatin; Wogonin.

Substrates: Apolipoproteins; Aspirin; Cholesterol; Lipids; Lipoproteins; LXR/PXR Ligands.

Inhibitors: 4,4'-Diisothiocyanatostilbene-2,2'-disulfonic acid disodium salt; Atorvastatin; Ezetimibe; Glyburide; GW 1929; miR-33; Naringenin; Phenytoin; Phosphatidylcholine; PPARG agonists; Rifampin; Simvastatin; Statins; Vistatin.

Inducers: ATI-111; Berberine; Charcoal-stripped fetal bovine serum; Cilostazol; GW 3965; Honokiol; Hormone Replacement Therapy; HT31; Ibuprofen; Low-density lipoproteins (LDL); Lycopene; Methotrexate; Methyl-3β-hydroxy-5α,6α-epoxycholestanol (MHEC); N-(2,2,2-Trifluoro-ethyl)-N-[4-(2,2,2-trifluoro-1-hydroxy-1-trifluoromethyl-ethyl)-phenyl]-benzenesulfonamide (TO-901317); N-3-Oxo-dodecanoyl-Homoserine lactone (3OC12-HSL); *Panax notoginseng* saponins; Phosphodiesterase (PDE) type 4 inhibitors; Phenylsulfone-substituted quinoxaline (WYE-672); PPARG agonists; Reginoid LG268; Ritonavir; Rolipram; Simvastatin; SIRT1; Statins; Stigmasterol; Tributyltin chloride (TBTC); Wogonin.

Animal Models

Humanized transgenic mouse line. Transgenic mice that express human *abca1* have been developed. Increased total *abca1* expression did not alter the pattern of *abca1* distribution, but resulted in increased cholesterol efflux, elevated HDL-C levels, and increased apoA1 and apoA2 expression. The *abca1* gene contains an internal promoter with LXR elements within intron 1. Activation of this functional internal promoter by oxysterols *in vivo* directly contributed to an increase in human-specific mRNA and protein levels.

Transgenic mice. Mice with a targeted inactivation of *abca1* display morphologic abnormalities and perturbations in their lipoprotein metabolism concordant with Tangier disease. *Abca1* is expressed on the plasma membrane and the Golgi complex, mediates apolipoprotein AI/APOA1-associated export of cholesterol and phospholipids from the cell, and is regulated by cholesterol flux. Structural and functional abnormalities in caveolar processing and the trans-Golgi secretory pathway of cells lacking functional ABC1 indicated that lipid export processes involving vesicular budding between the Golgi and the plasma membrane were severely disturbed. Lipid profiles in the knockout mice revealed a reduction of approximately 70% in cholesterol, markedly reduced plasma phospholipids, and an almost complete lack of high density lipoproteins, when compared with wild-type littermates. Dramatic alterations in HDL-C and near absence of APOA1 were found. Inactivation of the *abca1* gene led to an increase in the absorption of cholesterol in mice fed a chow or a high fat and high cholesterol diet. Histopathologic examination of knockout mice showed a striking accumulation of lipid-laden macrophages and type II pneumocytes in the lungs. The findings demonstrated that the knockout mice had pathophysiologic hallmarks of human Tangier disease and highlighted the capacity of *abca1* transporters to participate in the regulation of dietary cholesterol absorption.

Abca1 is expressed in Purkinje and cortical pyramidal neurons in the CNS, as well as in astrocytes and microglia. Astrocytes and microglia from *abca1*-null mice showed impaired ability to efflux cholesterol to exogenous apolipoprotein E (APOE), although residual efflux was present. The mutant cells showed increased intracellular lipid accumulation compared to wild-type cells. In addition, *Abca1*-null mice showed a 65% decrease in brain levels of APOE as a consequence of reduced secretion from mutant glial cells, with the hippocampus and striatum being the most severely affected brain regions. *Abca1* may play a role in cholesterol transport and APOE metabolism in the CNS. By analyzing brain tissue, cerebrospinal fluid, plasma, and primary astrocyte cultures from wild-type, *abca1*+/-, and *abca1*-/- mice, it was observed that deletion of *abca1* markedly affects metabolism of apoE and cholesterol in the CNS and in nascent lipoprotein particles secreted by cultured astrocytes.

Intestine-specific *abca1*-null mice were generated. Approximately 30% of the steady-state plasma HDL pool is contributed by intestinal *abca1* in mice. HDL derived from intestinal *abca1* appeared to be secreted directly into the circulation. Analysis of lymph from liver-specific *abca1*-null mice with very low plasma HDL showed that HDL in lymph was predominantly derived from the plasma compartment. Intestinal *abca1* may play a critical role in plasma HDL biogenesis *in vivo*. Mice with specific inactivation of *abca1* in pancreatic β-cells have been generated. In this model, markedly impaired glucose tolerance and defective insulin secretion but normal insulin sensitivity was observed. Islets isolated from these mice showed altered cholesterol homeostasis and impaired insulin secretion *in vivo*. Rosiglitazone, a thiazolidinedione, requires β-cell abca1 for its beneficial effects on glucose tolerance. Abca1 plays a role in β-cell cholesterol homeostasis and insulin secretion. Cholesterol accumulation may contribute to β-cell dysfunction in type 2 diabetes. 5' missense mutations were expressed at the plasma membrane but produced little or no apoA1-stimulated cholesterol efflux when transfected into HEK293 cells. All mutants except one showed a marked decline in interaction between the *abca1* mutant and apoA1. The deficits shown by these mutations establish their causality in Tangier disease, and binding of apoA1 by *abca1* is necessary, but not sufficient, to stimulate cholesterol efflux. 3 mutations in the first extracellular domain of *abca1* showed little or no apoA1-mediated HDL assembly when expressed in HEK293 cells. Two of these mutations were associated with impaired glycosylation, retention in the endoplasmic reticulum or the cis-Golgi complex, and failure to localize to the plasma membrane.

Allelic Variants

Total variants: 1532.

Selected SNPs

rs2230806. HGVS: NG_007981.1: g.74570G>A; NM_005502.2: c.656G>A; NP_005493.2: p.Arg219Lys; NT_008470.18: g.14942072C>T. Protection against coronary heart disease in familial hypercholesterolemia. In heterozygous familial hypercholesterolemia (FH) patients, the clinical expression of FH is highly variable in terms of the severity of hypercholesterolemia and the age at onset and severity of coronary heart disease (CHD). ABCA1 may play a key role in the onset of premature CHD in FH. The presence of the Arg219Lys variant in the *ABCA1* gene in FH patients influences the risk of CHD. The K allele of the R219K variant was significantly more frequent in FH patients without premature CHD than in those with premature CHD, suggesting that the genetic variant may influence development and progression of atherosclerosis in FH patients. The K allele of the R219K polymorphism seemed to modify CHD risk without important modification of plasma HDL-C levels, and it appeared to be more protective for smokers than nonsmokers. In a large Swedish population-based study of 1177 individuals with a first myocardial infarction event and 1526 healthy controls, an association between the R219K polymorphism and increased serum levels of apolipoprotein B (APOB) and LDL cholesterol was found among smokers, but not among nonsmokers. The *ABCA1*: Arg219Lys; K allele frequency differs markedly between blacks and whites, and the variant allele modulates the association between age and HDL cholesterol, as well as body fatness and triglycerides in a beneficial manner only in whites. Mutations in the *ABCA1* gene have been established as the molecular defect in Tangier disease and familial hypoalphalipoproteinemia, uncommon genetic disorders characterized by deficient or depressed high-density lipoprotein (HDL) cholesterol and increased triglycerides. Information regarding the frequency of common variants, including R219K within the coding region of the *ABCA1* gene and their effect on these phenotypes in the general population is limited. The frequency of the variant allele (K219) was higher in blacks than in whites (0.595 vs 0.262), with carriers (KK+RK) representing 83.8% of blacks vs 44.2% of whites. After adjusting for age, body mass index (BMI), and sex, the genotype effect on HDL-C and natural logarithm of triglycerides was not apparent in whites or blacks. However, significant interaction effects of genotype and age on HDL-C and genotype and BMI on triglycerides were found in whites. The *ABCA1* R219K polymorphism is associated with a higher HDL-C level in Asians and a protective role for CAD risk both in Asians and Caucasians.

Carriers (KK+RK), unlike noncarriers (RR) showed a positive relationship between age and HDL-C. Carriers of the KK genotype had higher levels of HDL-C than those of the RR genotype. Patients with the RR genotype had significantly higher serum triglyceride levels and lower HDL-C levels than those with the KK genotype. The effects of pravastatin in increasing HDL-C levels were significantly greater in patients with the KK genotype compared with those with the RR genotype. The R219K polymorphism was associated with altered lipoprotein levels and the R219K variant significantly modulated the HDL-C response to pravastatin in Chinese patients with CHD. Carriers of the G1051A allele were associated only with increased concentrations of HDL cholesterol.

ABCA1 polymorphisms were combined in haplotypes possessing G1051A, G2706A, G2868A and -565C/T; (AAGC) seemed to be most protective against significant stenosis, whereas GGAT was probably the most atherogenic. AAGC seems to be a protective haplotype whereas GGAT has an atherogenic effect in a Tunisian population.

Alleles: African: A: 0.688-0.797, G: 0.203-0.312; Asian: A: 0.398-0.467, G: 0.533-0.602; European: A: 0.158, G: 0.842; Multi-national: A: 0.250-0.487, G: 0.513-0.750; North American: A: 0.193-0.680, G: 0.320-0.807; Pacific: A: 0.521, G: 0.479. **Genotypes:** African: A/A: 0.417-0.444, A/G: 0.305-0.542, G/G: 0.041-0.051; Asian: A/A: 0.159-0.222, A/G: 0.477-0.489, G/G: 0.289-0.364; European: A/A: 0.017, A/G: 0.283, G/G: 0.700; Multi-national: A/A: 0.231-0.250, A/G: 0.000-0.513, G/G: 0.256-0.750; North American: A/A: 0.032, A/G: 0.323, G/G: 0.645; Pacific: A/A: 0.292, A/G: 0.458, G/G: 0.250.

rs3890182. HGVS: NG_007981.1: g.47782C>T; NM_005502.2: c.161-806C>T; NT_008470.18: g.14968860G>A. High density lipoprotein cholesterol level quantitative trait locus 13. Association of rs3890182 (74A-G) in the *ABCA1* gene with HDL-C levels. **Alleles:** African: A: 0.192, G: 0.808; Asian: A: 0.056-0.067, G: 0.933-0.944; European: A: 0.093, G: 0.907. **Genotypes:** African: A/A: 0.067, A/G: 0.250, G/G: 0.683; Asian: A/A: 0.000, A/G: 0.111-0.133, G/G: 0.867-0.889; European: A/A: 0.017, A/G: 0.153, G/G: 0.831.

rs3905000. HGVS: NG_007981.1: g.38367C>T; NM_005502.2: c.67-5594T>C; NT_008470.18: g.14978275G>A. Coronary heart disease. Risk allele: rs3905000-G. **Alleles:** African: A: 0.217-0.225, G: 0.775-0.783; Asian: A: 0.000-0.067, G: 0.933-1.000; European: A: 0.000-0.093, G: 0.907-1.000; Multi-national: A: 0.104, G: 0.896; North American: A: 0.090-0.167, G: 0.833-0.910. **Genotypes:** African: A/A: 0.050-0.067, A/G: 0.317-0.333, G/G: 0.617; Asian: A/A: 0.000, A/G: 0.000-0.133, G/G: 0.867-1.000; European: A/A: 0.000-0.017, A/G: 0.000-0.153, G/G: 0.831-1.000; Multi-national: A/A: 0.000, A/G: 0.208, G/G: 0.792; North American: A/A: 0.167, A/G: 0.000, G/G: 0.833.

rs1883025. HGVS: NG_007981.1: g.31136G>A; NM_005502.2: c.66+1594A>G; NT_008470.18: g.14985506C>T. Polygenic dyslipidemia. Risk allele: rs1883025-T. **Alleles:** African: A: 0.300-0.302, G: 0.698-0.700; Asian: A: 0.167-0.256, G: 0.744-0.833; European: A: 0.200-0.202, G: 0.798-0.800. **Genotypes:** African: A/A: 0.117-0.121, A/G: 0.362-0.367, G/G: 0.517; Asian: A/A: 0.000-0.044, A/G: 0.333-0.422, G/G: 0.533-0.667; European: A/A: 0.033-0.035, A/G: 0.333, G/G: 0.632-0.633.

rs28933692. HGVS: NG_007981.1: g.114326G>T; NM_005502.2: c.3295G>T; NP_005493.2: p.Asp1099Tyr; NT_008470.18: g.14902316C>A. High density lipoprotein deficiency. Cerebral amyloid angiopathy. A 3295G-T transversion, predicted to result in an Asp1099Tyr (D1099Y) mutation.

rs28937313. HGVS: NG_007981.1: g.110636A>G; NM_005502.2: c.2804A>G; NP_005493.2: p.Asn935Ser; NT_008470.18: g.14906006T>C. Tangier disease. A 3199A-G transition in exon 19 of the *ABCA1* gene, leading to an Asn935Ser missense mutation.

rs28937314. HGVS: NG_007981.1: g.110635A>C; NM_005502.2: c.2803A>C; NP_005493.2: p.Asn935His; NT_008470.18: g.14906007T>G. Tangier disease. A 3198A-C transversion in exon 19 of the *ABCA1* gene, resulting in an Asn935His missense mutation. This and the Asn935Ser mutation involved the Walker A motif of the first nucleotide-binding fold.

rs2066714. NG_007981.1: g.108684A>G; NM_005502.2: c.2649A>G; NP_005493.2: p.Le883Met; NT_008470.19: g.36751285T>C. Coronary artery disease. **Alleles:** African: A: 0.450-0.562, G: 0.438-0.550; Asian: A: 0.267-0.500, G: 0.500-0.733; European: A: 0.500-0.867, G: 0.133-0.500; Multi-national: A: 0.681-0.763, G: 0.237-0.319; North American: A: 0.520-0.903, G: 0.097-0.480; Pacific: A: 0.396, G: 0.604. **Genotypes:** African: A/A: 0.200-0.250, A/G: 0.500-0.625, G/G: 0.125-0.300; Asian: A/A: 0.089-0.273, A/G: 0.356-0.455, G/G: 0.273-0.556. European: A/A: 0.000-0.733, A/G: 0.267-1.000, G/G: 0.000; Multi-national: A/A: 0.490-0.632, A/G: 0.263-0.382, G/G: 0.105-0.128; North American: A/A: 0.000-0.806, A/G: 0.000-0.194, G/G: 0.000; Pacific: A/A: 0.167, A/G: 0.458, G/G: 0.375.

rs2066715. HGVS: NG_007981.1: g.107404G>A; NM_005502.2: c.2473G>A; NP_005493.2: p.Val825Ile; NT_008470.19: g.36752565C>T. Coronary artery disease. The subjects with AA genotype in Bai Ku Yao had higher serum TC levels than the subjects with GG and GA genotypes. V825I polymorphism is associated with male serum HDL-C and ApoA1 levels in the Han, and the serum TC levels in the Bai Ku Yao populations. **Alleles:** Multi-national: A: 0.013, G: 0.987. **Genotypes:** Multi-national: A/G: 0.026, G/G: 0.974.

rs2575879. HGVS: NC_000009.11: g.107668825G>C; NG_007981.1: g.26612C>G; NM_005502.3: c.-92-2773C>G. Associated with higher serum HDL-C concentrations.

rs1800978. HGVS: NC_000009.11: g.107665978C>G; NG_007981.1: g.29459G>C; NM_005502.3: c.-18G>C. Associated with lower HDL-C concentrations.

rs9282541. HGVS: NC_000009.11: g.107620835G>A; NG_007981.1: g.74602C>T; NM_005502.3: c.688C>T; NP_005493.2: p.Arg230Cys. In individuals with the R230C/C230C genotypes, lower HDL-C levels were observed compared to those with the R230R genotype. The C230 allele was associated with an increased risk for hypoalphalipoproteinemia. The non-synonymous Arg230Cys variant of *ABCA1* is associated with low levels of HDL cholesterol levels in Mexican adults. The variant R230C, apparently exclusive to Native American individuals, was associated with low HDL-C levels, obesity and type 2 diabetes in Mexican Mestizos. The C230 allele was found in 29 of 36 Native American groups, but not in European, Asian or African individuals.

c.5398A>C and c.2369G>A: Associated with HDL cholesterol deficiency in serum.

110-bp Ins/14-bp Del: Tangier disease. An insertion of a 110-bp DNA fragment structurally related to the Alu sequence family of repetitive sequences and deletion of 14 bp in exon 12 of the *ABCA1* gene. This insertion/deletion predicted deletion of 6 aa and an in-frame insertion of 38 residues. This mutation does not allow the synthesis of the normal ABC1 transporter.

1-bp Del, 1764G: Tangier disease. A 1-bp deletion, removing guanine at nucleotide 1764. This mutation, localized in codon 548, created a frameshift that led to a premature translation stop 26 aa downstream of the deletion site. The translation product was predicted to be nonfunctional because it lacked 75% of the amino acid sequence, including all transmembrane regions and both ATP-binding cassettes.

1-bp Del, 2665C: Tangier disease.

2-bp Del, 3283TC: Tangier disease. A dinucleotide deletion in exon 22: 3283-3284TC. The deletion resulted in a frameshift mutation and a premature stop codon starting at position 3375. The gene product was predicted to encode a nonfunctional protein of 1084 aa, which is approximately half the size of the full-length ABC1 protein.

3-bp Del: High density lipoprotein deficiency, type 2. A 3-bp deletion that resulted in loss of nucleotides 2017-2019 and deletion of a leucine at position 633, which is conserved in mouse and *C. elegans*.

4-bp Del, 3787CGCC: Familial high density lipoprotein deficiency. A 4-bp deletion (CGCC) at nucleotide 3787, resulting in premature termination by frameshift at codon 1224.

Ala877Val: Tangier disease. A2750C-T transition, changing alanine to valine (missense mutation).

Arg1680Trp: Tangier disease, variant. An Arg1680Trp missense mutation.

Arg2021Trp: Tangier disease. A 6181C-T transition in exon 47, resulting in an Arg2021Trp (R2021W) substitution.

Asn875Ser: Tangier disease. A 2744A-G transition that changes asparagine to serine in the highly conserved Walker A motif of the amino terminal ATP-binding fold.

Asp1229Asn: Tangier disease. A 3805G-A transition in exon 27 of the *ABCA1* gene, resulting in an Asp1229Asn change.

Cys1417Arg: Tangier disease. T-to-C transition in exon 30 resulting in a Cys1417Arg substitution.

Gln537Arg: Tangier disease. An A-to-G transition at nucleotide 1730 in exon 13, resulting in the substitution of arginine for a conserved glutamine at residue 537.

Int12-14 Del, Int16-31 Del: Tangier disease. A double deletion in the *ABCA1* gene: 1.2 kb from intron 12-14 and 19.9 kb from intron 16-31, which encodes the sixth transmembrane region (a linker region) and the seventh transmembrane region of the putative secondary structure. The 3'-deletion junction had an insertion of 21 bp. The 16 bp within the 21-bp insertion was not found in the original sequence, but was complementary to the proximal sequence of the 5'-deletion junction. The same oriented Alu sequence was found in both intron 14 and intron 31, facilitating the stabilization of the folding of the *ABCA1* gene to promote nonhomologous intragenic recombination. Double deletions in the same gene previously reported for dystrophin; in the beta-globin gene (*HBB*); in the growth hormone gene (*GH1*); and in the *GALNS* gene, which is mutant in mucopolysaccharidosis type IVA. A simultaneous event of double deletions was proposed for the case of thalassemia patients with changes in the *HEB* gene due to inversion between deletions.

Ivs24DS, G-C: Tangier disease. A G-to-C transversion in the splice donor site of exon 24, predicted to cause alternative splicing, deleting a significant part of the transcript.

Phe2009Ser: High density lipoprotein deficiency. Cerebral amyloid angiopathy. A 5966T-C transition, predicted to result in a Phe2009Ser mutation.

Ser1446Leu: Tangier disease. Ser1446Leu missense amino acid substitution.

Trp530Ser: Tangier disease. A 1709G-C transversion, resulting in a Trp530Ser aa substitution (missense mutation).

Tyr573Ter: Tangier disease. A 2033C-A transversion in exon 12 of the *ABCA1* gene, resulting in conversion of codon 573 from TAC (Tyr) to TAA (Ter)(Y573X).

Pro85Leu: Low HDL-C levels.

Arg1851Gln: Low HDL-C levels.

Arg1925Gln: Scott syndrome.

Ivs46: Del T-39-46: Low HDL-C levels.

The variants: **219K** (rs2230806), **883I** (rs2066714), and **1587R** (rs2230808), or promoter variant (**14T**) in concert with the *APOE-4* allele, increase the risk of Alzheimer's disease.

-14C>T: Associated with high baseline gene-expression levels.

C69T: Influences CHD risk and response to statin treatment. The TT genotype was associated with 1.7 times higher CHD risk than the CC genotype.

R1587K (rs2230808-KK) and **R219K** (rs2230806-RK) polymorphisms were associated with significantly increased small HDL. The R1587KKK genotype and the R219K KK genotype were also associated with increased HDL particle concentrations.

Other selected SNPs

rs1800978. HGVS: NG_007981.1: g.4728G>C; NT_008470.18: g.15011914C>G. **Frequency:** C: 0.485, G: 0.515.

rs2246298. HGVS: NG_007981.1: g.4704C>T; NT_008470.18: g.15011938G>A. **Frequency:** C: 0.804, T: 0.196.

rs2740483. HGVS: NG_007981.1: g.4902C>G; NT_008470.18: g.15011740G>C. **Frequency:** C: 0.253, G: 0.747.

rs56064613. HGVS: NG_007981.1: g.4896C>T; NT_008470.18: g.15011746G>A. **Frequency:** C: 0.989, T: 0.011.

rs4149338. HGVS: NG_007981.1: g.149534C>T; NM_005502.2: c.*693C>T; NT_008470.18: g.14867108G>A. **Alleles:** African: C: 0.610, T: 0.390; Asian: C: 0.193-0.300, T: 0.700-0.807; European: C: 0.716, T: 0.284; Multi-national: C: 0.530, T: 0.470. **Genotypes:** African: C/C: 0.373, C/T: 0.475, T/T: 0.153; Asian: C/C: 0.045-0.111, C/T: 0.295-0.378, T/T: 0.511-0.659; European: C/C: 0.552, C/T: 0.328, T/T: 0.121.

rs13306080. HGVS: NG_007981.1: g.149209A>G; NM_005502.2: c.*368A>G; NT_008470.18: g.14867433T>C. **Alleles:** African: A: 1.000, G: 0.000; Asian: A: 0.989-1.000, G: 0.000-0.011; European: A: 1.000, G: 0.000. **Genotypes:** African: A/A: 1.000, A/G: 0.000; Asian: A/A: 0.978-1.000, A/G: 0.000-0.022; European: A/A: 1.000, A/G: 0.000.

rs10991377. HGVS: NG_007981.1: g.152061A>G; NM_005502.2: c.*3220A>G; NT_008470.18: g.14864581T>C.

rs34879708. HGVS: NG_007981.1: g.148784C>A; NM_005502.2: c.6729C>A; NP_005493.2: p.Asp2243Glu; NT_008470.18: g.14867858G>T. **Alleles:** North American: G: 0.987, T: 0.013. **Genotypes:** North American: G/G: 0.974, G/T: 0.026.

rs363717. HGVS: NG_007981.1: g.150373G>A; NM_005502.2: c.*1896G>A; NT_008470.18: g.14865905C>T. Associated with thalidomide-related peripheral neuropathy. **Alleles:** African: A: 0.933, G: 0.067; Asian: A: 0.740-0.833, G: 0.167-0.260; European: A: 0.793-0.847, G: 0.153-0.207; Multi-national: A: 0.820-0.880, G: 0.120-0.180; North American: A: 0.790-0.933, G: 0.067-0.210; Pacific: A: 0.780-0.800, G: 0.200-0.220. **Genotypes:** African: A/A: 0.867, A/G: 0.133, G/G: 0.000; Asian: A/A: 0.700, A/G: 0.267, G/G: 0.033; European: A/A: 0.586-0.699, A/G: 0.295-0.414, G/G: 0.000-0.006; North American: A/A: 0.867, A/G: 0.133, G/G: 0.000; Pacific: A/A: 0.640, A/G: 0.280-0.320, G/G: 0.040-0.080.

rs1800977. HGVS: NG_007981.1: g.4987C>T; NT_008470.18: g.15011655G>A. **Alleles:** African: C: 0.800, T: 0.200; Asian: C: 0.678-0.682, T: 0.318-0.322; European: C: 0.642, T: 0.358; Multi-national: C: 0.630, T: 0.370; Other: C: 0.766, T: 0.234. **Genotypes:** African: C/C: 0.633, C/T: 0.333, T/T: 0.033; Asian: C/C: 0.444-0.500, C/T: 0.364-0.467, T/T: 0.089-0.136; European: C/C: 0.450, C/T: 0.383, T/T: 0.167.

rs2230808. HGVS: NG_007981.1: g.132633A>G; NM_005502.2: c.4760A>G; NP_005493.2: p.Lys1587Arg; NT_008470.18: g.14884009T>C. **Alleles:** African: A: 0.709-0.883, G: 0.117-0.292; Asian: A: 0.289-0.389, G: 0.611-0.711; European: A: 0.158-0.161, G: 0.839-0.842; Multi-national: A: 0.200-0.473, G: 0.527-0.800; North American: A: 0.261-0.850, G: 0.150-0.739; Pacific: A: 0.417, G: 0.583. **Genotypes:** African: A/A: 0.500-0.783, A/G: 0.200-0.417, G/G: 0.017-0.083; Asian: A/A: 0.111-0.178, A/G: 0.356-0.422, G/G: 0.400-0.533; European: A/A: 0.033-0.034, A/G: 0.250-0.254, G/G: 0.712-0.717; Multi-national: A/A: 0.200-0.405, A/G: 0.000-0.135, G/G: 0.459-0.800; North American: A/A: 0.087, A/G: 0.348, G/G: 0.565; Pacific: A/A: 0.167, A/G: 0.500, G/G: 0.333.

rs4149339. HGVS: NG_007981.1: g.150281C>T; NM_005502.2: c.*1440C>T; NT_008470.18: g.14866361G>A. **Alleles:** African: C: 0.608-0.610, T: 0.390-0.392; Asian: C: 0.186-0.300, T: 0.700-0.814; European: C: 0.717-0.724, T: 0.276-0.283; Multi-national: C: 0.580, T: 0.420. **Genotypes:** African: C/C: 0.367-0.373, C/T: 0.475-0.483, T/T: 0.150-0.153; Asian: C/C: 0.047-0.111, C/T: 0.279-0.378, T/T: 0.511-0.674; European: C/C: 0.550-0.552, C/T: 0.333-0.345, T/T: 0.103-0.117.

rs4149340. HGVS: NG_007981.1: g.150752C>T; NM_005502.2: c.*1911C>T; NT_008470.18: g.14865890G>A. **Alleles:** African: C: 0.925, T: 0.075; Asian: C: 0.800-0.852, T: 0.148-0.200; European: C: 1.000, T: 0.000; Multi-national: C: 0.960, T: 0.040. **Genotypes:** African: C/C: 0.850, C/T: 0.150, T/T: 0.000; Asian: C/C: 0.622-0.705, C/T: 0.295-0.356, T/T: 0.000-0.022; European: C/C: 1.000, C/T: 0.000, T/T: 0.000.

rs2482432. HGVS: NG_007981.1: g.152265A>G; NT_008470.18: g.14864377T>C. **Alleles:** African: C: 0.200, T: 0.800; Asian: C: 0.057-0.078, T: 0.922-0.943; European: C: 0.425, T: 0.575. **Genotypes:** African: C/C: 0.017, C/T: 0.367, T/T: 0.617; Asian: C/C: 0.000-0.023, C/T: 0.068-0.156, T/T: 0.844-0.909; European: C/C: 0.200, C/T: 0.450, T/T: 0.350.

rs2066718. HGVS: NG_007981.1: g.106182G>A; NM_005502.2: c.2311G>A; NP_005493.2: p.Val771Met; NT_008470.18: g.14910460C>T. **Alleles:** African: A: 0.075, G: 0.925; Asian: A: 0.022-0.057, G: 0.943-0.978; European: A: 0.000, G: 1.000; Multi-national: A: 0.051-0.070, G: 0.930-0.949. **Genotypes:** African: A/G: 0.150, G/G: 0.850; Asian: A/G: 0.044-0.114, G/G: 0.886-0.956; European: A/G: 0.000, G/G: 1.000; Multi-national: A/G: 0.103, G/G: 0.897.

rs9282543. HGVS: NG_007981.1: g.96061T>C; NM_005502.2: c.1196T>C; NP_005493.2: p.Val399Ala; NT_008470.18: g.14920581A>G. **Alleles:** African: C: 0.000, T: 1.000; Asian: C: 0.000, T: 1.000; European: C: 0.009, T: 0.991; Multi-national: C: 0.010, T: 0.090; North American: C: 0.000-0.033, T: 0.967-1.000; Pacific: C: 0.000, T: 1.000. **Genotypes:** African: C/T: 0.000, T/T: 1.000; Asian: C/T: 0.000, T/T: 1.000; European: C/T: 0.017, T/T: 0.983; Multi-national: C/T: 0.020, T/T: 0.980; North American: C/T: 0.000-0.065, T/T: 0.935-1.000; Pacific: C/T: 0.000, T/T: 1.000.

Evolution

The *ABCA1* and *ABCA2* genes map to 9q22-q31 and 9q34, respectively. In the mouse, the homologs map to chromosomes 4 and 2, respectively, in regions showing homology of synteny to human 9q. The ancestral chromosome split in the mouse lineage at an evolutionary breakpoint situated between hexabrachion and gelsolin, both of which map to human chromosome 9 and to mouse chromosomes 4 and 2, respectively. Thus, *ABCA1* and *ABCA2* probably originated through a duplication event that took place before speciation and predated the splitting of the ancestral chromosome equivalent to human 9q. Their degree of sequence similarity, less impressive than that of the P-glycoprotein isoforms, also argues for a duplication event occurring at an earlier evolutionary stage.

Genomics and Pharmacogenomics

ABCA1 and ABCA2 mutant phenotypes: Cholesterol homeostasis in the enterocyte is regulated by the interplay of multiple genes that ultimately determines the net amount of cholesterol reaching the circulation from the small intestine. The effect of deleting these genes, particularly acyl CoA:cholesterol acyl transferase 2 (*ACAT2*), on cholesterol absorption and fecal sterol excretion is well documented. The intestinal mRNA level for adenosine triphosphate-binding cassette transporter A1 (*ABCA1*) increases in *Acata*^{-/-} mice. Turley et al measured the expression of numerous genes and related metabolic parameters in the intestine and liver of *ACAT2*-deficient mice fed diets containing either added cholesterol or ezetimibe, a selective sterol absorption inhibitor. Cholesterol feeding raised the concentration of UC in the small intestine, and this was accompanied by a significant reduction in the relative mRNA level for Niemann-Pick C1-like 1 (*NPC1L1*) and an increase in the mRNA level for both *ABCA1* and *ABCG5/8*. All these changes were reversed by ezetimibe. When mice deficient in both *ACAT2* and *ABCA1* were fed a high-cholesterol diet, the increase in intestinal UC levels was no greater than it was in mice lacking only *ACAT2*. This resulted from a combination of compensatory mechanisms including diminished *NPC1L1*-mediated cholesterol uptake, increased cholesterol efflux via *ABCG5/8*, and possibly rapid cell turnover.

ABCA7: ATP-binding cassette transporter (ABC) A7 is an ABC family protein that is a so-called full-size ABC transporter, highly homologous to *ABCA1*, which mediates the biogenesis of high-density lipoprotein (HDL) with cellular lipid and helical apolipoproteins. *ABCA7* mediates the formation of HDL when exogenously transfected and expressed; however, endogenous *ABCA7* was shown to have no significant impact on the generation of HDL and was found to be associated with phagocytosis regulated by sterol regulatory element binding protein 2. Since phagocytosis is one of the fundamental functions of animal cells as an important responsive reaction to infection, injury and apoptosis, *ABCA7* seems to be one of the key molecules linking sterol homeostasis and the host defense system. According to data reported by Tanaka et al, HDL apolipoproteins were shown to enhance phagocytosis by stabilizing *ABCA7* against calpain-mediated degradation and increasing its activity, shedding light on a new aspect of the regulation of the host-defense system.

The ATP-binding cassette transporter (ABC)A7 strongly associates with phagocytic function of, rather than biogenesis of high-density lipoprotein (HDL), being regulated by sterol regulatory element binding protein (SREBP)2. Phagocytic activity was found enhanced by apolipoprotein (apo)A-I and apoA-II more than twice the maximum in J774 and mouse peritoneal macrophages. Tanaka et al investigated the molecular basis of this reaction in association with the function of *ABCA7*. Similar to *ABCA1*, *ABCA7* was degraded, probably by calpain, and apoA-I and apoA-II stabilized *ABCA7* against degradation. Cell surface biotinylation experiments demonstrated that endogenous *ABCA7* predominantly resides on the cell surface and that the apolipoproteins increase the surface *ABCA7*. The increase of phagocytosis by apolipoproteins was retained in the J774 cells treated with *ABCA1* siRNA and in the peritoneal macrophages from *ABCA1*-knockout mice, but it was abolished in the J774 cells treated with *ABCA7* siRNA and in the peritoneal macrophages from *ABCA7*-knockout mice. Phagocytosis was decreased in the cells in the peritoneal cavity of the *ABCA7*-knockout mouse compared with the wild-type control. Extracellular helical apolipoproteins augment *ABCA7*-associated phagocytosis by stabilizing *ABCA7*.

ABCG1: ATP binding cassette transporter G1 (*ABCG1*) mediates the transport of cellular cholesterol to HDL, and it plays a key role in maintaining macrophage cholesterol homeostasis. During inflammation, HDL undergoes substantial remodeling, acquiring lipid changes and serum amyloid A (SAA) as a major apolipoprotein. de Beer et al investigated whether remodeling of HDL that occurs during acute inflammation impacts *ABCG1*-dependent efflux. Lipid-free SAA acts similarly to apolipoprotein A-I (apoA-I) in mediating sequential efflux from *ABCA1* and *ABCG1*. Compared with normal mouse HDL, acute phase (AP) mouse HDL containing SAA exhibited a modest but significant 17% increase in *ABCG1*-dependent efflux. Interestingly, AP HDL isolated from mice lacking SAA (SAKO mice) was even more effective in promoting *ABCG1* efflux. Hydrolysis with Group IIA secretory phospholipase A₂ (sPLA₂-IIA) significantly reduced the ability of AP HDL from SAAKO mice to serve as a substrate for *ABCG1*-mediated cholesterol transfer, indicating that phospholipid (PL) enrichment, and not the presence of SAA, is responsible for alterations in efflux. AP human HDL, which is not PL-enriched, was somewhat less effective in mediating *ABCG1*-dependent efflux compared with normal human HDL. Inflammatory remodeling of HDL impacts *ABCG1*-dependent efflux independent of SAA.

Endothelin-1 (ET-1), a potent proatherogenic vasoconstrictive peptide, is known to promote macrophage foam cell formation via mechanisms that are not fully understood. Excessive lipid accumulation in macrophages is a major hallmark during the early stages of atherosclerotic lesions. Cholesterol homeostasis is tightly regulated by scavenger receptors (SRs) and ATP-binding cassette (ABC) transporters during the transformation of macrophage foam cells. Lin et al investigated the possible mechanisms by which ET-1 affects lipid accumulation in macrophages. Oxidized low density lipoprotein (oxLDL) treatment increases lipid accumulation in rat bone marrow-derived macrophages. Combined treatment with ET-1 and oxLDL significantly exacerbated lipid accumulation in macrophages as compared to treatment with oxLDL alone. ET-1 markedly decreased the *ABCG1* levels via ET type A and B receptors and activation of the phosphatidylinositol 3-kinase pathway; however, ET-1 had no effect on the protein expression of CD36, SR-BI, SR-A, or *ABCA1*. ET-1 treatment did not affect *ABCG1* mRNA expression. ET-1 decreases *ABCG1* possibly due to the enhancement of the proteasome/calpain pathway-dependent degradation of *ABCG1*. ET-1 significantly reduced the efficiency of the cholesterol efflux in macrophages. ET-1 may impair cholesterol efflux and further exacerbate lipid accumulation during the transformation of macrophage foam cells.

Lipid and cholesterol metabolism in the postprandial phase is associated with both quantitative and qualitative remodeling of HDL particle subspecies that may influence their anti-atherogenic functions in the reverse cholesterol transport pathway. Julia et al evaluated the capacity of whole plasma or isolated HDL particles to mediate cellular free cholesterol (FC) efflux, cholesterol transfer protein (CETP)-mediated cholesteryl ester (CE) transfer, and selective hepatic CE uptake during the postprandial phase in subjects displaying type IIb hyperlipidemia. Postprandial, large HDL2 displayed an enhanced capacity to mediate FC efflux via both scavenger receptor class B type I (SR-BI)-dependent (+12%) and ATP binding cassette transporter G1 (*ABCG1*)-dependent (+31%) pathways *in vitro* cell systems. The capacity of whole postprandial plasma (4 h and 8 h postprandially) to mediate cellular FC efflux via the *ABCA1*-dependent pathway was significantly increased (+19%). Concomitantly, postprandial lipemia was associated with elevated endogenous CE transfer rates from HDL2 to apoB lipoproteins and with attenuated capacity (-17%) of total HDL to deliver CE to hepatic cells. Postprandial lipemia enhanced SR-BI and *ABCG1*-dependent efflux to large HDL2 particles. Postprandial lipemia is equally associated with deleterious features by enhancing formation of CE-enriched, triglyceride-rich lipoprotein particles through the action of CETP and by reducing the direct return of HDL-CE to the liver.

Adiponectin: Expression of the activator protein (AP) 2 β is closely associated with type-2 diabetes and negatively regulates expression of the adiponectin gene. Iwamoto and Yokoyama reported that AP-2 α negatively regulates the ATP-binding cassette A1 (*ABCA1*) gene through its Ser-phosphorylation by protein kinase (PK) D. The phosphorylation site of AP-2 α located in the basic domain, a critical site for its DNA binding, is conserved among species and five subtypes of AP-2. Knockdown of *PKD* by its siRNA led to the increase in the mRNA and the promoter activity of adiponectin, and resulted in increase of adiponectin secretion and decrease of fat accumulation in cultured cells. PKD activators decreased expression of the adiponectin gene, and its inhibition by *PKD* siRNA and by a selective inhibitor G06983 canceled this effect. ChIP analysis demonstrated that inhibition of PKD activity decreased the DNA binding of AP-2 β to the adiponectin promoter. *PKD* is a common modulator of the DNA binding activity of AP-2 α and AP-2 β through their phosphorylation for negative regulation of the *ABCA1* and adiponectin genes expression, respectively.

alpha-Lipoic acid: α -Lipoic acid (α -LA), a key cofactor in cellular energy metabolism, has protective activities in atherosclerosis. Cheng et al examined whether α -LA affects foam cell formation and its underlying molecular mechanisms in murine macrophages. Treatment with α -LA markedly attenuated oxidized low-density lipoprotein (oxLDL)-mediated cholesterol accumulation in macrophages, which was due to increased cholesterol efflux. Additionally, α -LA treatment

dose-dependently increased protein levels of ATP-binding cassette transporter A1 (ABCA1) and ABCG1 but had no effect on the protein expression of SR-A, CD36, or SR-BI involved in cholesterol homeostasis. α -LA increased the mRNA expression of ABCA1 and ABCG1. α -LA-mediated suppression of oxLDL-induced lipid accumulation was abolished by GGPP or LXR α siRNA treatment. LXR α -dependent upregulation of ABCA1 and ABCG1 may mediate the beneficial effect of α -LA on foam cell formation.

Alzheimer's disease: Variant (219K, 883I, and 1587R), or promoter variant (14T) in concert with the *APOE-4* allele, increase the risk of Alzheimer's disease. Both single marker alleles and haplotypes of ABCA1 contribute to variable cerebrospinal fluid tau and β -amyloid protein levels, and brain β -amyloid load. Results indicate that variants of ABCA1 may affect the risk of Alzheimer's disease (AD), providing support for a genetic link between AD and cholesterol metabolism. In 42 individuals with AD, an association was found between increased CSF cholesterol and β -amyloid protein levels. In a separate study of a Swedish population of 1177 individuals with a first myocardial infarction event and 1526 controls, an association between the R219K polymorphism and serum levels of apolipoprotein B (APOB) and LDL-C was found among smokers, but not among nonsmokers.

Aberrant cholesterol metabolism has been implicated in Alzheimer's disease (AD). Underexpression of NPC1 in concert with under-expression of ABCA1 would result in increased cholesterol accumulation and increased AD risk. Rodríguez-Rodríguez examined a functional polymorphism in the ABCA1 promoter region (-477, rs2422493), and four NPC1 polymorphisms in exon 6 (rs18050810), intron 20 (rs4800488), intron 22 (rs2236707), and intron 24 (rs2510344) capturing 85% of genetic variability in the Hap Map Caucasian (CEU) population, in a group of 631 Spanish AD patients and 731 controls. Subjects carrying both the ABCA1 (-477) TT genotype and the NPC1 (exon 6) GG genotype, NPC1 (intron 20) AA genotype, NPC1 (intron 22) AA genotype or NPC1 (intron 24) GG genotype may have higher risk of developing AD than subjects without these risk genotypes.

ATP-binding cassette transporter A1 (ABCA1) reduces amyloid-beta burden in transgenic mouse models of AD. Associations between ABCA1 polymorphisms and AD risk are also established. Kim et al assessed ABCA1 mRNA and protein expression in the hippocampus of AD cases compared to controls. ABCA1 was clearly expressed in hippocampal neurons and expression was increased two- to three-fold in AD cases. The increased hippocampal ABCA1 expression was associated with increased APOE and PUMA gene expression, implying an association with neuronal stress. Consistent with this, treatment of SK-N-SH neurons with amyloid-beta peptide resulted in a 48% loss in survival and a significant upregulation of ABCA1, APOE, and PUMA gene expression. Studies in young (2 month) and old (12 month) transgenic mice expressing a familial AD form of human amyloid-beta protein precursor and presenilin-1 revealed a significant age-dependent upregulation of hippocampal ABCA1 compared to wild-type control mice. However, hippocampal APOE and Puma gene expression were not correlated with increased Abca1 expression in mice. ABCA1 is upregulated in AD hippocampal neurons potentially via an amyloid-beta-mediated pathway.

Cholesterol content of cerebral membranes is tightly regulated by elaborate mechanisms that balance the level of cholesterol synthesis, uptake and efflux. Among the conventional regulatory elements, a recent research focus has been nuclear receptors, a superfamily of ligand-activated transcription factors providing an indispensable regulatory framework in controlling cholesterol metabolism pathway genes. The mechanism of transcriptional regulation by nuclear receptors such as LXRs involves formation of heterodimers with RXRs. LXR/RXR functions as a sensor of cellular cholesterol concentration and mediates cholesterol efflux by inducing the transcription of key cholesterol shuffling vehicles, namely ATP-binding cassette transporter A1 (ABCA1) and ApoE. Akram et al provided evidence from direct analysis of human *post-mortem* brain gene and protein expression suggesting that RXR α , a key regulator of cholesterol metabolism, is differentially expressed in individuals with dementia. RXR expression showed strong association with ABCA1 and ApoE gene expression, particularly in Alzheimer's disease vulnerable regions. LXR/RXR-induced upregulation of ABCA1 and ApoE levels may be the molecular determinants of cholesterol dyshomeostasis and of the accompanying dementia observed in Alzheimer's disease.

ApoE-1 mimetic peptide D4-F: Adenosine triphosphate-binding cassette transporter A1 (ABCA1) plays a crucial role in apolipoprotein A-I (apoA-I) binding activity and promotes cellular cholesterol efflux. ApoA-I mimetic peptide D4-F has been reported to have similar ability to apoA-I. Liu et al investigated the effects of D4-F on ABCA1 expression and ABCA1-dependent cholesterol efflux and examined the role of the Cdc42/cyclic adenosine monophosphate (cAMP)/protein kinase A (PKA) pathway on the regulation of ABCA1 by D4-F in THP-1 macrophage-derived foam cells. D4-F stabilized ABCA1 protein and enhanced ABCA1-dependent cholesterol efflux but had no effect on ABCA1 messenger RNA expression. D4-F enhanced cAMP level and PKA activity and ABCA1 serine phosphorylation. Short interfering RNA of PKA led to reduction of ABCA1 serine phosphorylation and ABCA1-mediated cholesterol efflux compensated by D4-F. PKA-specific activation by PKA agonist enhanced the upregulation of ABCA1 serine phosphorylation and ABCA1-mediated cholesterol efflux by D4-F. ABCA1 expression did not change by treatment with PKA agonist or PKA-short interfering RNA. Scramine B, a Cdc42 inhibitor, reduced the cAMP level compensated by D4-F. D4-F enhances ABCA1 serine phosphorylation and ABCA1-dependent cholesterol efflux through the Cdc42/cAMP/PKA pathway in THP-1 macrophage-derived foam cells.

ARL7: Ligand activation of liver X receptors (LXRs) has been shown to impact both lipid metabolism and inflammation. One complicating factor in studies utilizing synthetic LXR agonists is the potential for pharmacologic and receptor-independent effects. Hong et al described an LXR gain-of-function system that does not depend on the addition of exogenous ligand. They generated transgenic mice expressing a constitutively active VP16-LXR α protein from the *ap2* promoter. These mice exhibit increased LXR signaling selectively in adipose tissue and macrophages. Analysis of gene expression in primary macrophages derived from two independent VP16-LXR α transgenic lines confirmed the ability of LXR to drive expression of genes involved in cholesterol efflux and fatty acid synthesis. VP16-LXR α expression also suppressed the induction of inflammatory genes by lipopolysaccharide to a degree comparable to that of a synthetic agonist. They further utilized VP16-LXR α -expressing macrophages to identify and validate new targets for LXRs, including the gene encoding ADP-ribosylation factor-like 7 (ARL7). ARL7 has previously been shown to transport cholesterol to the membrane for ABCA1-associated removal and thus may be integral to the LXR-dependent efflux pathway. The ARL7 promoter contains a functional LXRE and can be transactivated by LXRs in a sequence-specific manner, indicating that ARL7 is a direct target of LXR.

Arsenic: Padovani et al set out to determine whether arsenic inhibits transcriptional activation of the liver X receptor (LXR)/retinoid X receptor (RXR) heterodimers, thereby impairing cholesterol efflux from macrophages and potentially contributing to a proatherogenic phenotype. Arsenic is an important environmental contaminant and has been linked to an increased incidence of atherosclerosis. Arsenic inhibits transcriptional activation of type 2 nuclear receptors, known to

heterodimerize with RXR. Environmentally relevant arsenic doses decrease the LXR/RXR ligand-induced expression of the LXR target genes (*ABCA1* and *SREBP-1c*). Arsenic failed to decrease cAMP-induced ABCA1 expression, suggesting a selective LXR/RXR effect. This selectivity correlated with the ability of arsenic to decrease LXR/RXR ligand-induced, but not cAMP-induced, cholesterol efflux. Arsenic inhibits the ability of LXR/RXR ligands to induce activation markers on the ABCA1 and *SREBP-1c* promoters and blocks ligand-induced release of the nuclear receptor coexpressor (NCoR) from the promoter. Arsenic did not alter the ability of LXR to transrepress inflammatory gene transcription, further supporting the hypothesis that RXR is the target for arsenic inhibition. Exposure to arsenic enhances the risk of atherosclerosis. Arsenic may promote an atherosclerotic environment by decreasing the ability of macrophages to efflux excess cholesterol, thereby favoring increased plaque formation.

Aspirin: The efflux of cellular cholesterol mediated by apolipoprotein (apo)A-I and ATP-binding cassette transporter A1 (ABCA1) is a major pathway of reverse cholesterol transport. Wang et al investigated the effect of aspirin on this process. Aspirin increased apoA-I-mediated cholesterol efflux and increased the expression of ABCA1. By increasing the dose of aspirin, ABCA1 expression and function were significantly decreased. In cells transfected with a specific peroxisome proliferator-activated receptor (PPAR)- α small interfering RNA, the induction of ABCA1 expression and apoA-I-mediated ³H-cholesterol efflux by aspirin were substantially suppressed. Low-dose aspirin increases ABCA1 expression via a PPAR- α -dependent mechanism and increases apoA-I-mediated cholesterol efflux.

Atherosclerosis: ABCA1 protein degradation is regulated by a PEST sequence (a sequence rich in proline, glutamic acid, serine, and threonine) in ABCA1 and is mediated by calpain protease. In a novel form of positive feedback control, the interaction of ABCA1 with APOA1 leads to inhibition of calpain protease degradation and an increase in ABCA1 protein on the cell surface. ABCA1 degradation by calpain may represent a novel therapeutic approach to increasing macrophage cholesterol efflux and decreasing atherosclerosis.

Insulin resistance/hyperinsulinism is one of the major risks for atherosclerotic vascular diseases. Low HDL may be involved in the pathogenesis of atherosclerosis. Nonomura et al examined the effects of insulin on HDL biosynthesis, focusing on the activity of ATP-binding cassette transporter A1 (ABCA1) in culture cells and in experimental animals. Insulin impairs HDL biosynthesis through modulation of ABCA1 activity by two different mechanisms. Insulin enhances degradation of ABCA1. This effect was found due to phosphorylation of ABCA1, which leads to a decrease in its specific activity. The authors identified a novel insulin-specific phosphorylation site of ABCA1, Tyr1206, which regulates its specific activity.

A crucial step in atherogenesis is the infiltration of the subendothelial space of large arteries by monocytes where they differentiate into macrophages and transform into lipid-loaded foam cells. Macrophages are heterogeneous cells that adapt their response to environmental cytokines. Th1 cytokines promote monocyte differentiation into M1 macrophages, whereas Th2 cytokines trigger an "alternative" M2 phenotype. Chinetti-Gbaguidi et al reported the presence of CD68⁺ mannose receptor (MR⁺) M2 macrophages in human atherosclerotic plaques. Histological analysis revealed that CD68⁺ MR⁺ macrophages locate far from the lipid core of the plaque and contain smaller lipid droplets compared to CD68⁺ MR⁻ macrophages. Interleukin (IL)-4-polarized CD68⁺ MR⁺ macrophages display a reduced capacity to handle and efflux cellular cholesterol because of low expression levels of the nuclear receptor liver x receptor (LXR α) and its target genes, ABCA1 and apolipoprotein E, attributable to the high 15-lipoxygenase activity in CD68⁺ MR⁺ macrophages. By contrast, CD68⁺ MR⁻ macrophages highly express opsonins and receptors involved in phagocytosis, resulting in high phagocytic activity. In M2 macrophages, peroxisome proliferator-activated receptor (PPAR) γ activation enhances the phagocytic but not the cholesterol trafficking pathways. These data reflect the existence of a distinct macrophage subpopulation with a low susceptibility to become foam cells but high phagocytic activity resulting from different regulatory activities of the PPAR γ -LXR α pathways.

Atherosclerosis is an inflammatory disease characterized by the accumulation of macrophages in the arterial intima. The activated macrophages secrete more proinflammatory cytokines, such as tumor necrosis factor (TNF)- α , which promote the development of the disease. Apolipoprotein A1 (apoA1), the major component of high-density lipoprotein (HDL), is involved in reverse cholesterol transport (RCT) of lipid metabolism. ApoA1 suppresses inflammation via repression of inflammatory cytokine expression. Yin et al reported that apoA1 suppresses the expression of some inflammatory cytokines induced by lipopolysaccharide (LPS) via a specific post-transcriptional regulation process, namely mRNA destabilization, in macrophages. AU-rich elements (AREs) in the 3' untranslated region (UTR) of TNF- α mRNA are responsive to the apoA1-mediated mRNA destabilization. The apoA1-induced inflammatory cytokine mRNA destabilization was associated with increased expression of mRNA-destabilizing protein tristetraprolin (TTP) through a JAK2/STAT3 signaling pathway dependent manner. When blocking interaction of apoA1 with ATP-binding membrane cassette transporter A1 (ABCA1), a major receptor for apoA1 in macrophages, it almost totally abolished the effect of apoA1 on TTP expression.

Macrophages cannot limit the uptake of lipids and rely on cholesterol efflux mechanisms for maintaining cellular cholesterol homeostasis. Important mediators of macrophage cholesterol efflux are ATP-binding cassette transporter 1 (ABCA1), which mediates the efflux of cholesterol to lipid-poor apolipoprotein A1, and scavenger receptor class B type I (SR-BI), which promotes efflux to mature high-density lipoprotein. Zhao et al studied the putative synergistic roles of ABCA1 and SR-BI in foam cell formation and atherosclerosis. Low-density lipoprotein receptor knockout (LDLR KO) mice were transplanted with bone marrow from ABCA1/SR-BI double knockout mice, the respective single knockouts, or wild-type littermates. Serum cholesterol levels were lower in ABCA1/SR-BI double knockout transplanted animals, as compared to the single knockout and wild-type transplanted animals on Western-type diet. Despite the lower serum cholesterol levels, massive foam cell formation was found in macrophages from spleen and the peritoneal cavity. BI double knockout transplanted animals also showed a major increase in proinflammatory KC (murine interleukin-8) and interleukin-12p40 levels in the circulation. After 10 weeks of Western-type diet feeding, atherosclerotic lesion development in the aortic root was more extensive in the LDLR KO mice reconstituted with ABCA1/SR-BI double knockout bone marrow. Deletion of ABCA1 and SR-BI in bone marrow-derived cells enhances *in vivo* macrophage foam cell formation and atherosclerotic lesion development in LDLR KO mice on Western diet, indicating that under high dietary lipid conditions, both macrophage ABCA1 and SR-BI contribute significantly to cholesterol homeostasis in the macrophage *in vivo* and are essential for reducing the risk for atherosclerosis.

Atherosclerosis has been characterized as a chronic inflammatory response to cholesterol deposition in arteries. Plasma high density lipoprotein (HDL) levels bear a strong independent inverse relationship with atherosclerotic cardiovascular disease. One central antiatherogenic role of HDL is believed to be its ability to remove excessive peripheral cholesterol back to the liver for subsequent catabolism and excretion, a physiologic process termed reverse cholesterol transport (RCT). Cholesterol efflux from macrophage foam cells, the initial step of RCT, is the most relevant step with respect to atherosclerosis. The ATP-binding cassette (ABC) transporters ABCA1 and

Pharmacogenomic Synopsis

Drug	Related Genes	Database	Protein	Inducer
Abacavir				
Anti-Infection Agent: Antiretroviral, Nucleoside and Nucleotide Reverse Transcriptase Inhibitor (ATC Code: J05AF06)	ACV1: ACV1001, ACV1002, ACV1003, ACV1004, ACV1005, ACV1006, ACV1007, ACV1008, ACV1009, ACV1010, ACV1011, ACV1012, ACV1013, ACV1014, ACV1015, ACV1016, ACV1017, ACV1018, ACV1019, ACV1020, ACV1021, ACV1022, ACV1023, ACV1024, ACV1025, ACV1026, ACV1027, ACV1028, ACV1029, ACV1030, ACV1031, ACV1032, ACV1033, ACV1034, ACV1035, ACV1036, ACV1037, ACV1038, ACV1039, ACV1040, ACV1041, ACV1042, ACV1043, ACV1044, ACV1045, ACV1046, ACV1047, ACV1048, ACV1049, ACV1050, ACV1051, ACV1052, ACV1053, ACV1054, ACV1055, ACV1056, ACV1057, ACV1058, ACV1059, ACV1060, ACV1061, ACV1062, ACV1063, ACV1064, ACV1065, ACV1066, ACV1067, ACV1068, ACV1069, ACV1070, ACV1071, ACV1072, ACV1073, ACV1074, ACV1075, ACV1076, ACV1077, ACV1078, ACV1079, ACV1080, ACV1081, ACV1082, ACV1083, ACV1084, ACV1085, ACV1086, ACV1087, ACV1088, ACV1089, ACV1090, ACV1091, ACV1092, ACV1093, ACV1094, ACV1095, ACV1096, ACV1097, 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